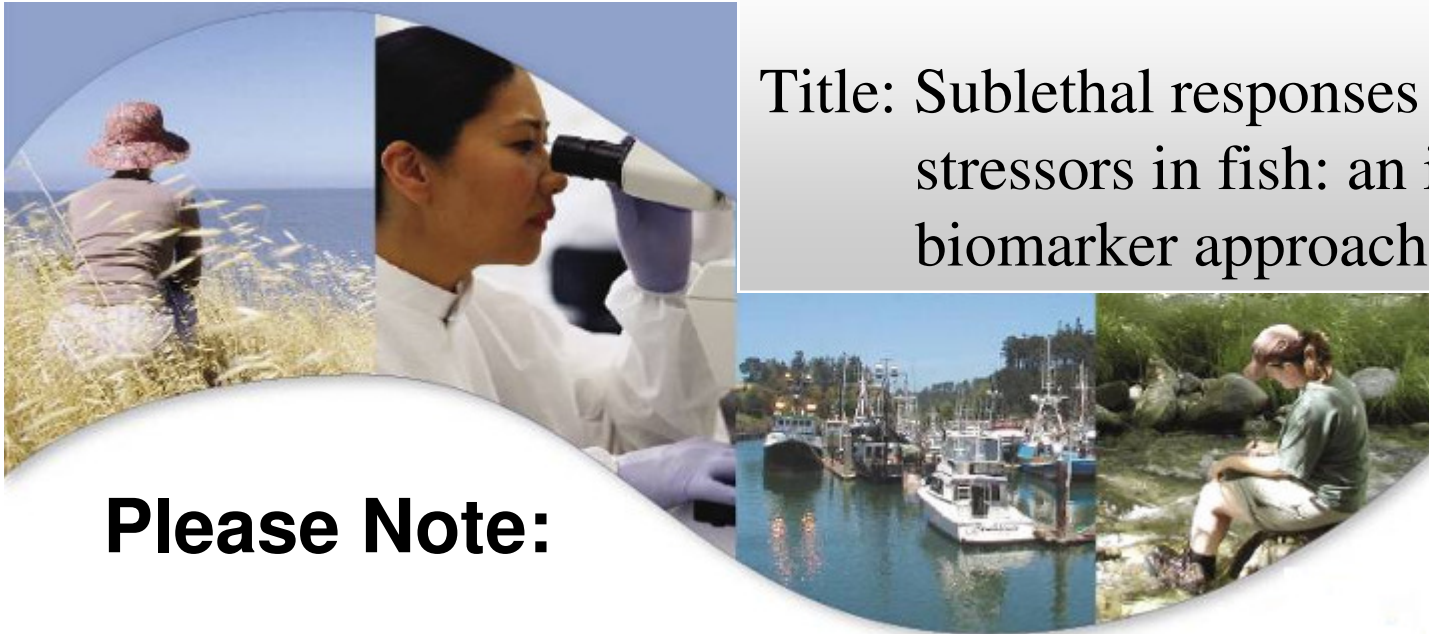


Welcome to SWAMP Webinar

Title: Sublethal responses to environmental stressors in fish: an integrative multi-biomarker approach



**Please Note:**

- All participants are muted upon log in
- For questions, all participants will be unmuted after the presentation is completed
- Put yourself on mute by pressing \*6 so that you do not detract other participants



Aquatic Toxicology Laboratory  
School of Veterinary Medicine - UC Davis

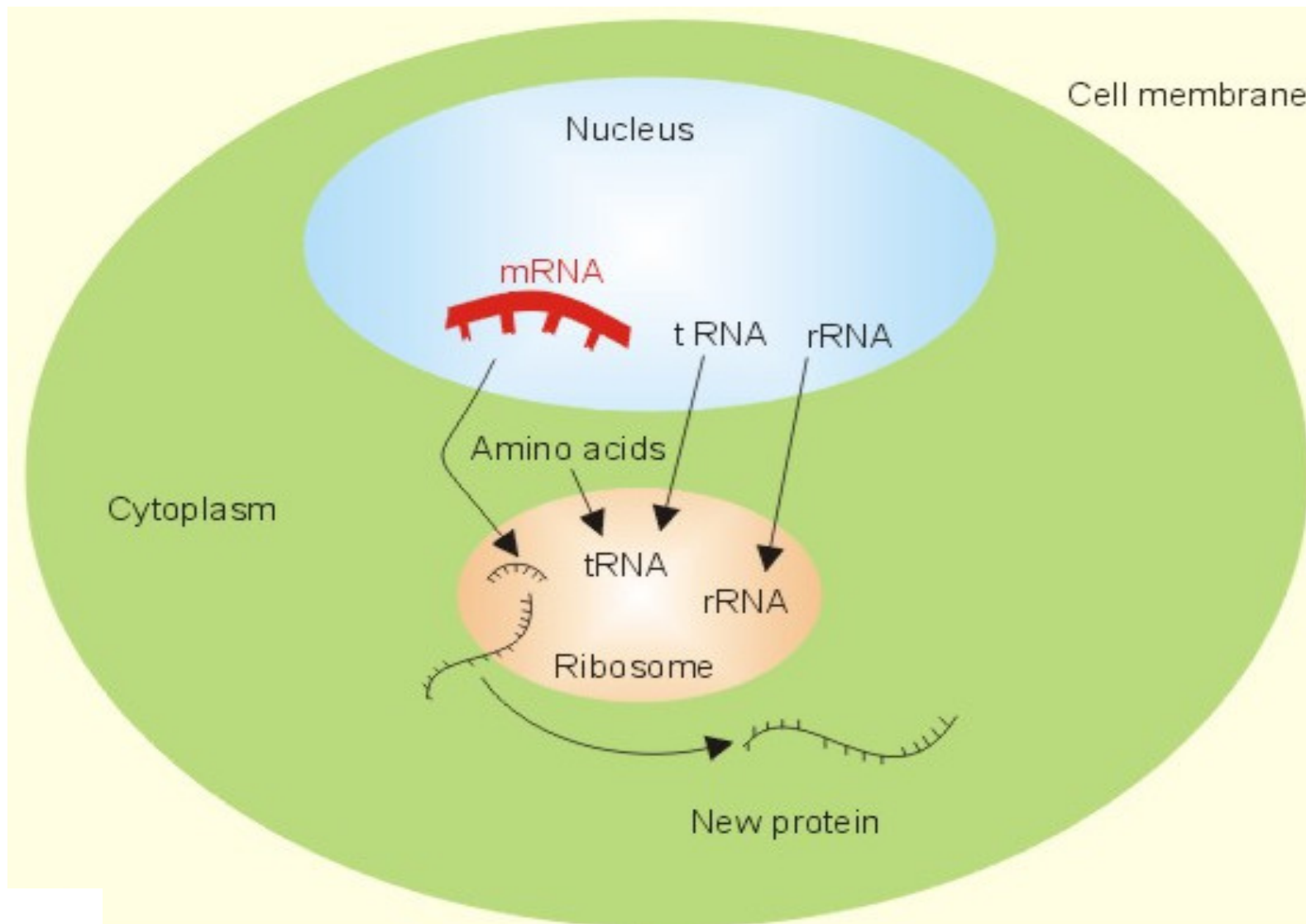


# Sublethal responses to environmental stressors in fish: an integrative multi-biomarker approach

Richard Connon, Sebastian Beggel, Erika Fritsch,  
Leandro D'Abronzo, Becca Hudson-Davies,  
Linda Deanovic, and Inge Werner

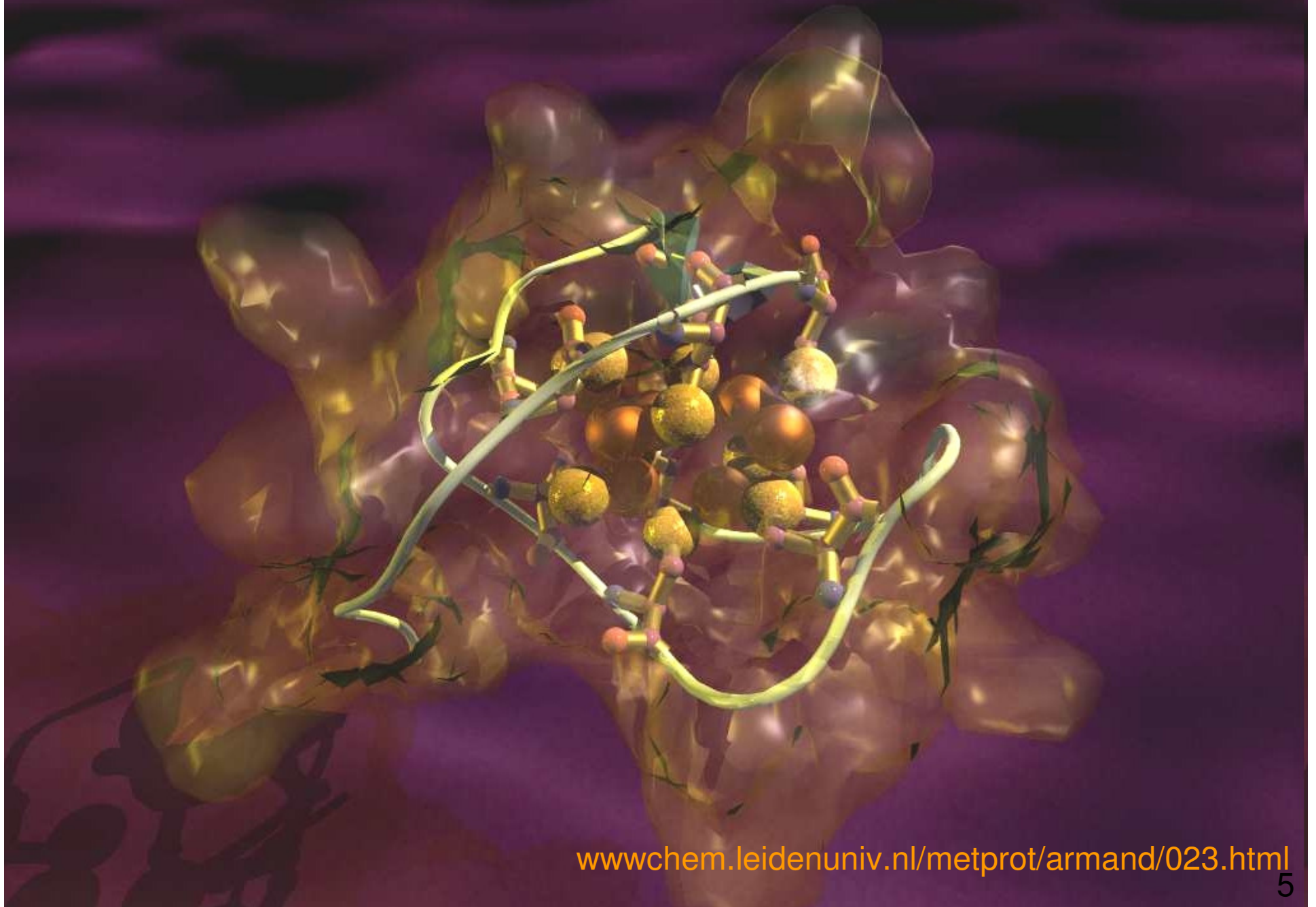
# Outline

- Molecular techniques background (5'-10')
- Links to relevant higher levels of organization
- Application of multiple biomarkers in delta smelt
- Ammonia/um in river water study
- Current, future and proposed work
- Other studies in model species (Inge Werner)



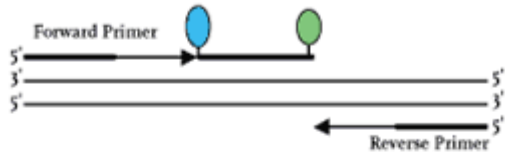


e.g. Metallothionein

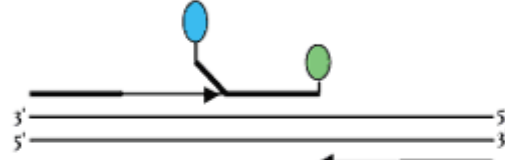


[wwwchem.leidenuniv.nl/metprot/armand/023.html](http://wwwchem.leidenuniv.nl/metprot/armand/023.html)

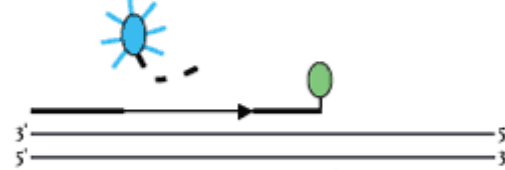
### Polymerization



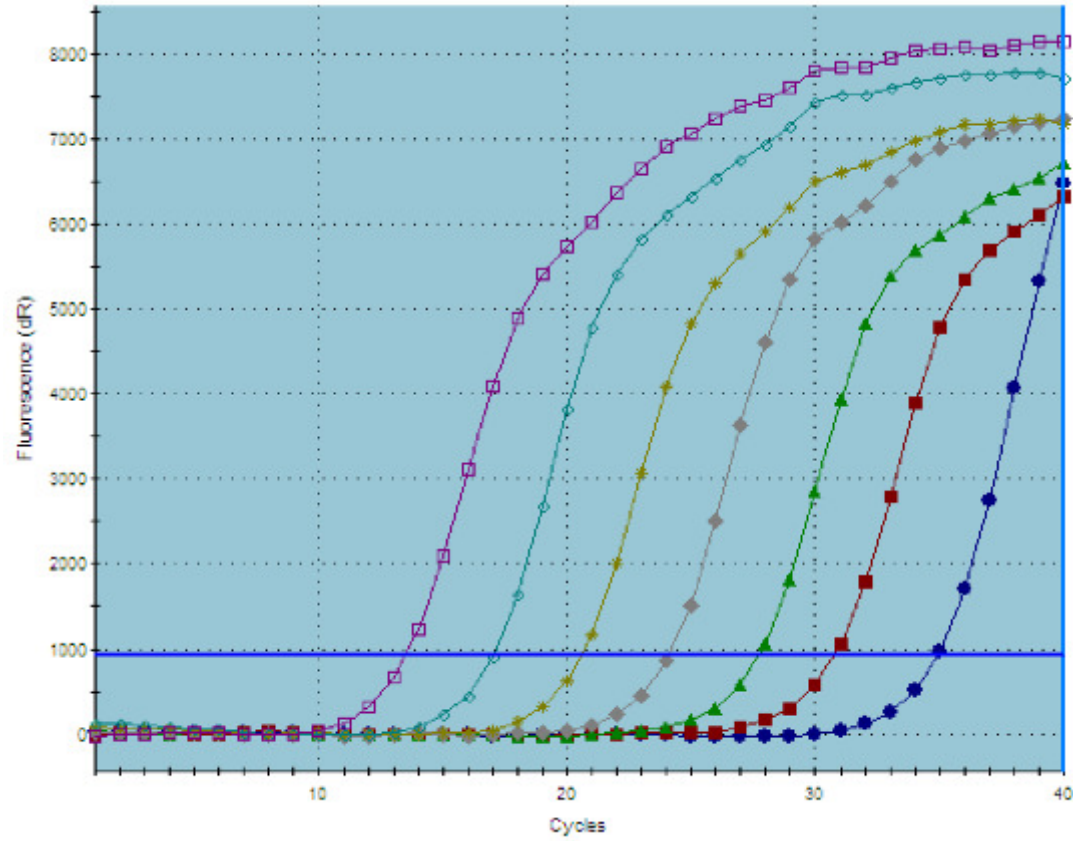
### Strand Displacement



### Cleavage



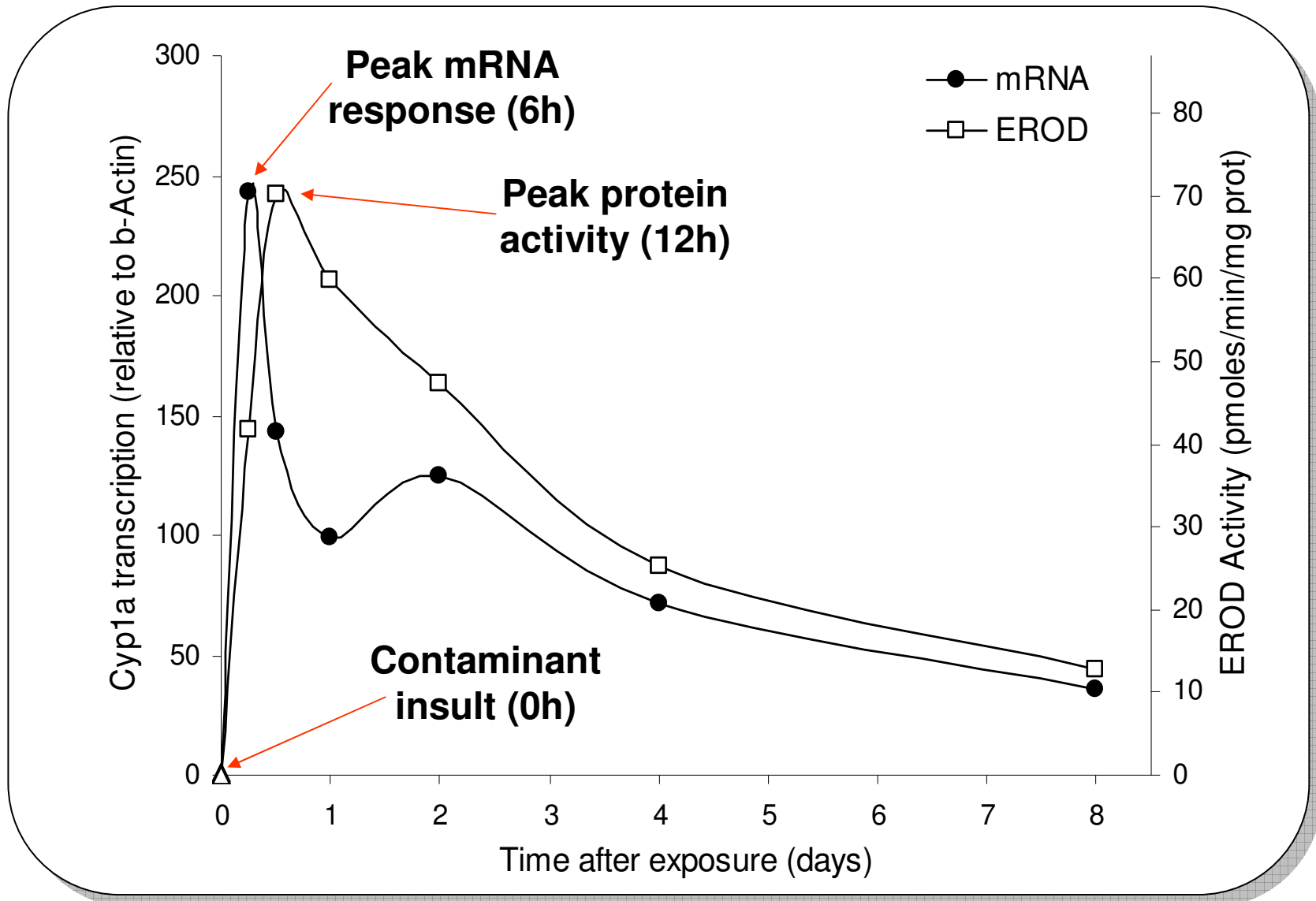
### Polymerization Completed



# Quantitative PCR

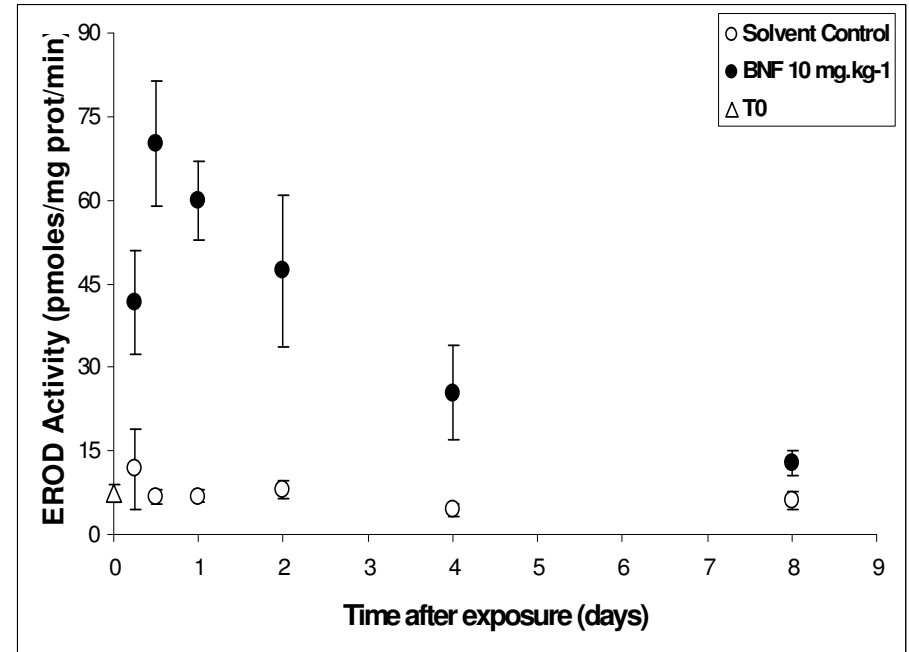
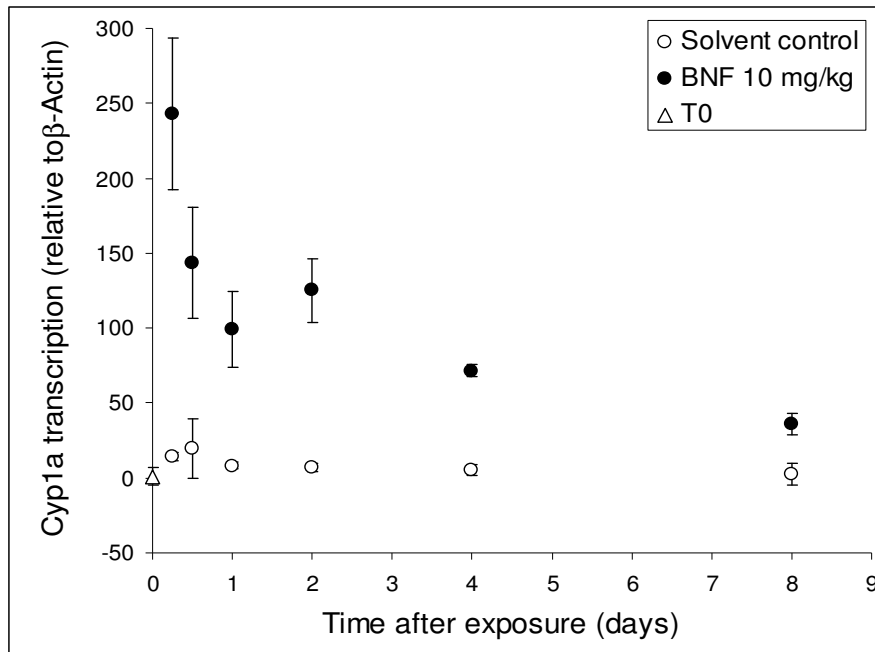


Connon RE, Durieux EDH, D'Abronzio LS, Ostrach DJ, and Werner I. Cytochrome P450 induction in the striped bass (*Morone saxatilis*): signature biomarkers of past and present xenobiotic exposure (in prep).



Time-dependent change in CYP1A **mRNA transcription** and **protein activity** (EROD) expression in striped bass exposed to  $10\text{mg.kg}^{-1}$   $\beta$ -naphthoflavone. 7

Connon RE, Durieux EDH, D'Abronzio LS, Ostrach DJ, and Werner I. Cytochrome P450 induction in the striped bass (*Morone saxatilis*): signature biomarkers of past and present xenobiotic exposure (in prep).

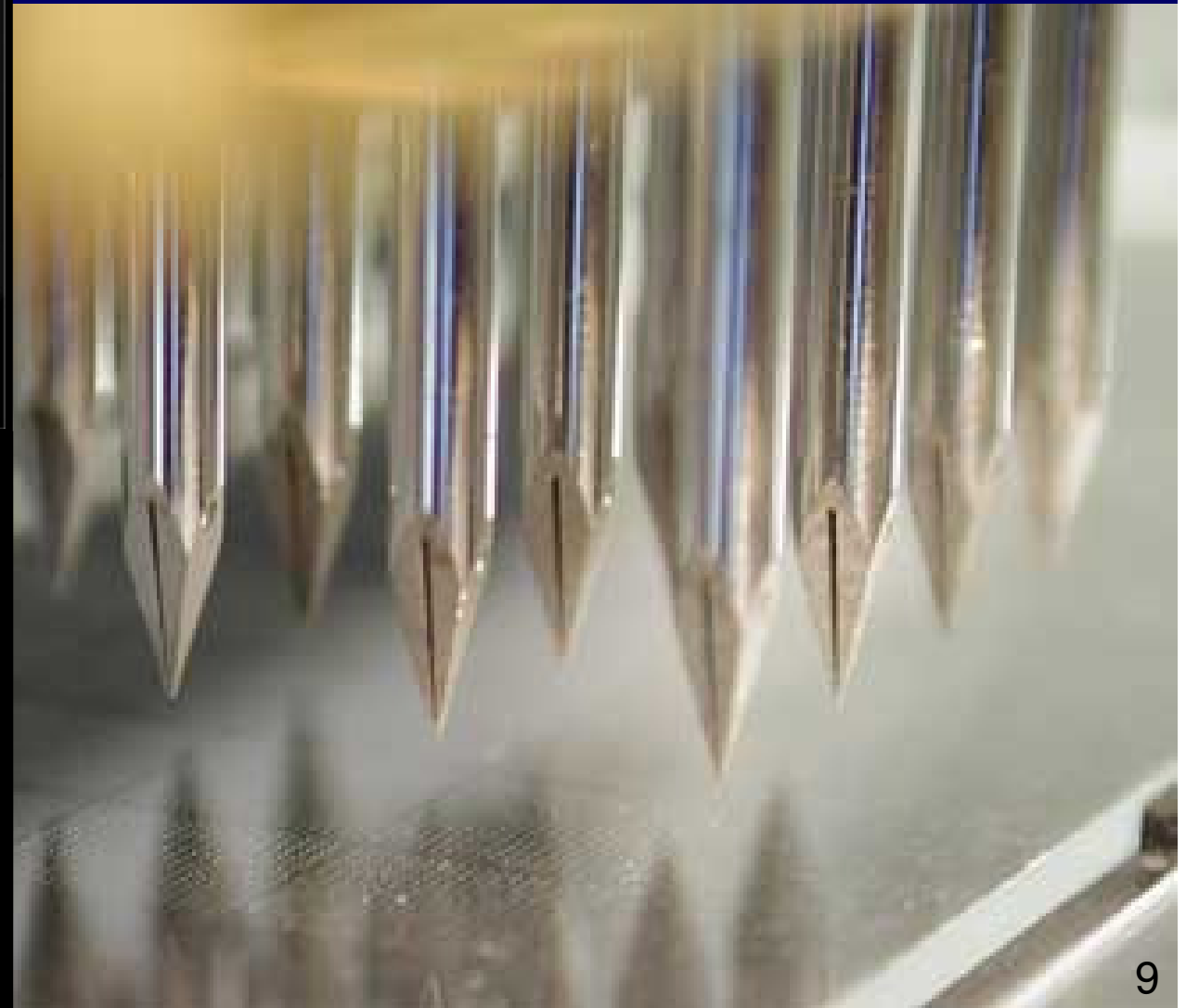
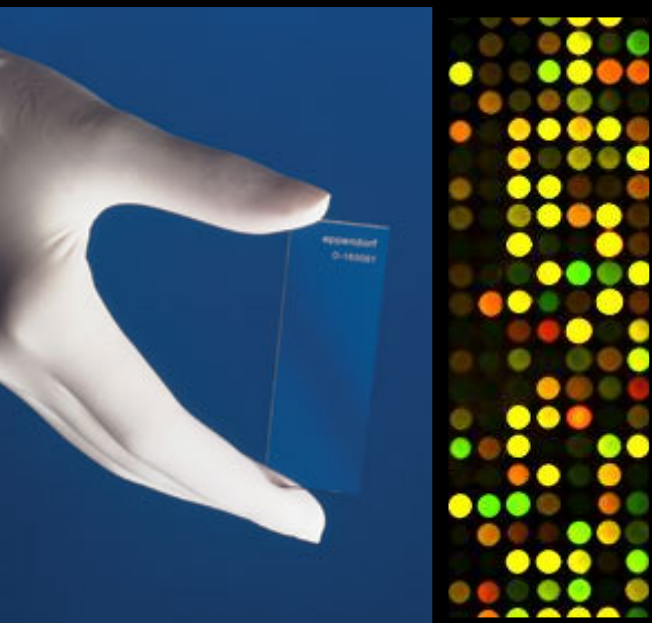
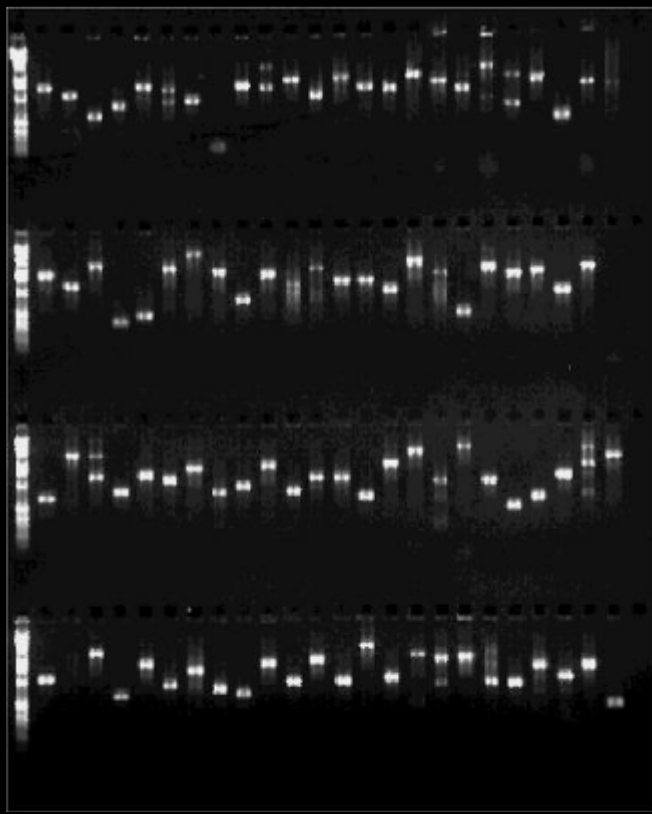


Time-dependent change in CYP1A **mRNA transcription** and **protein activity** (EROD) expression in striped bass exposed to 10mg.kg<sup>-1</sup>  $\beta$ -naphthoflavone. 8

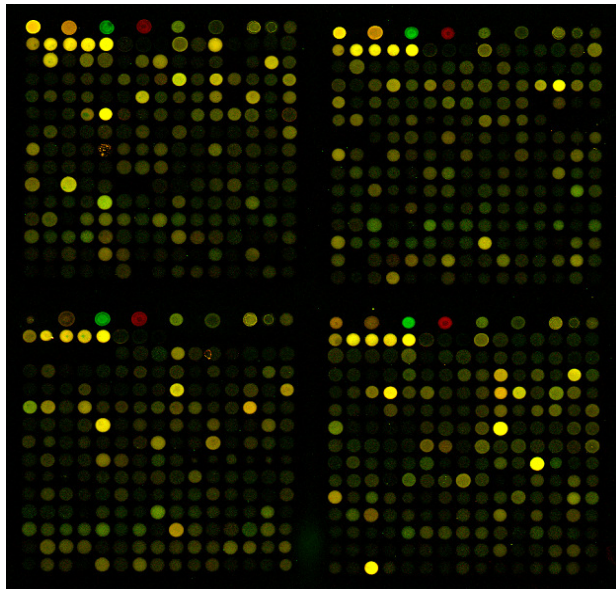


# Microarrays

e.g. cDNA library, PCR amplified fragments, purified, and printed onto epoxysilane coated glass slides.

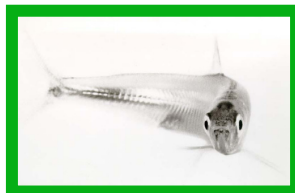


# What is a microarray and how does it work?

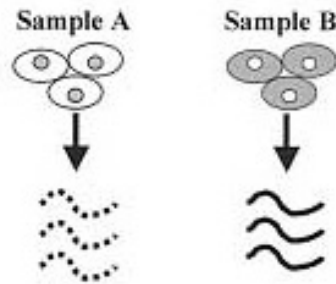


Exposed

Control

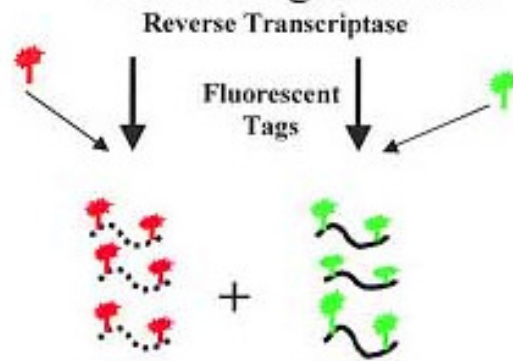


## A. RNA Isolation

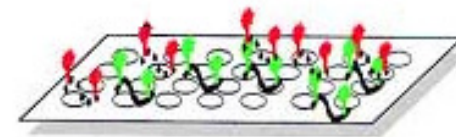
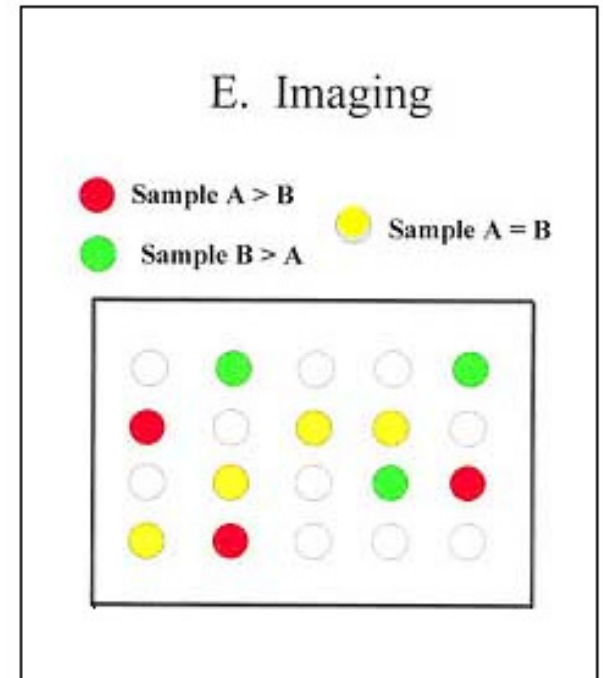


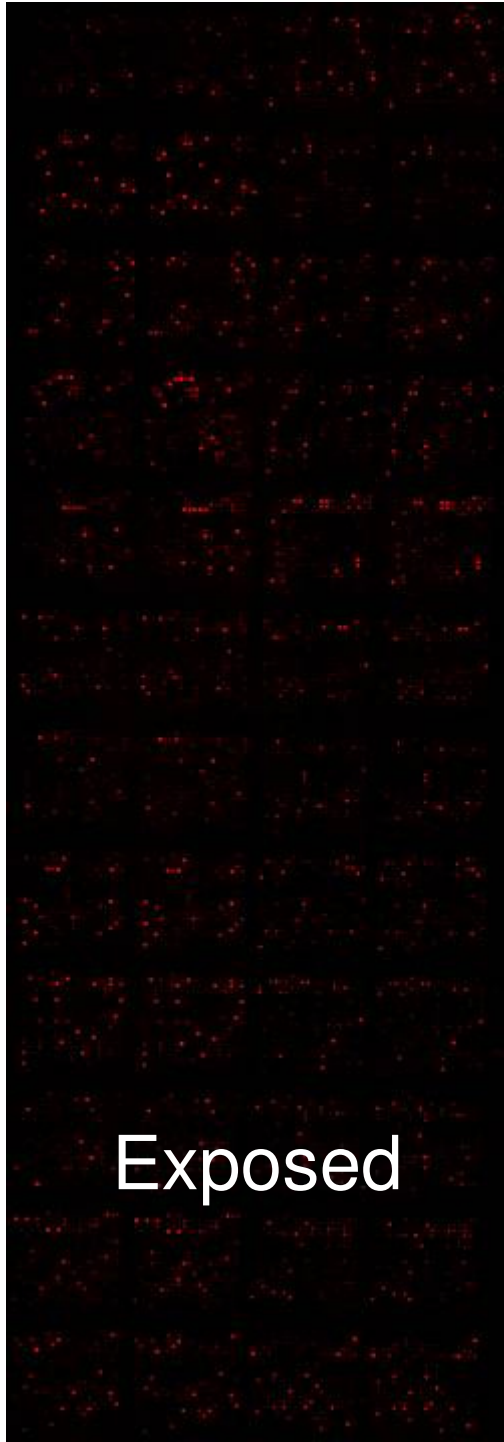
## B. cDNA Generation

## C. Labeling of Probe

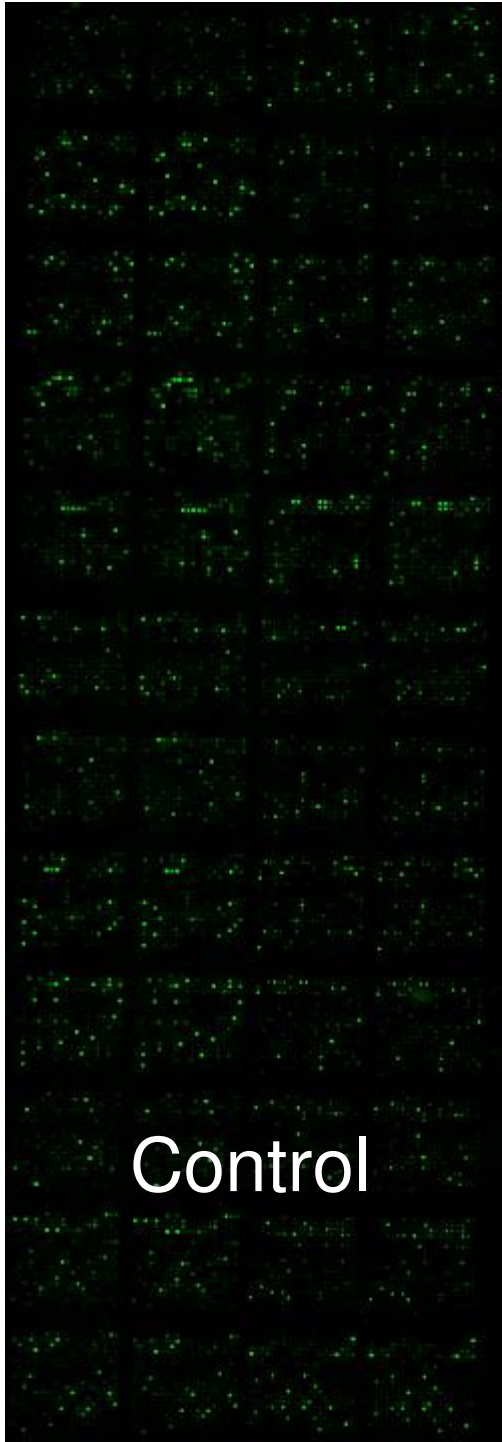


## D. Hybridization to Array

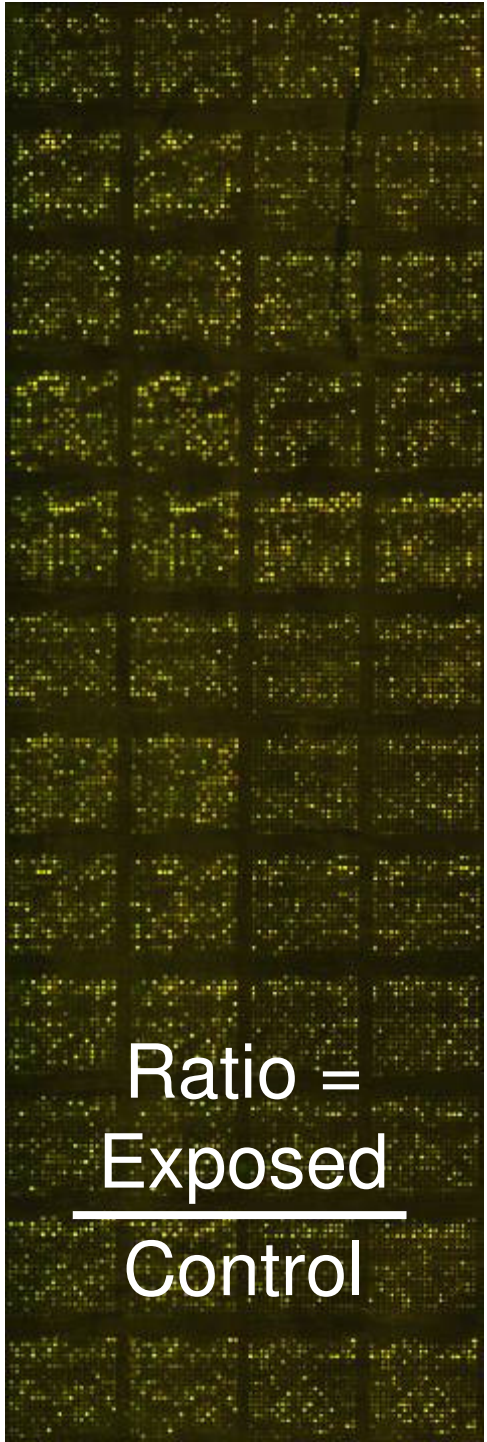




/



=

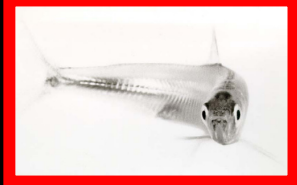




Smelt Clone ID	Response	Fold	Gene most similar to	Species Match	Accession No	E-Value
DS[96]_39_B_06	Up	2.36	1-acylglycerol-3-phosphate O-acyltransferase 3	<i>Danio rerio</i>	NP_998590	4.00E-68
DS[96]_77_C_03	Up	4.88	actin alpha 2, skeletal muscle	<i>Pagrus major</i>	BAF80060	1.00E-94
DS[96]_03_E_12	Up	3.75	actin, alpha 2, smooth muscle, aorta	<i>Danio rerio</i>	AAH75896	e-107
DS[96]_19_B_07	Up	2.92	actin, alpha, cardiac muscle 1 like	<i>Danio rerio</i>	NP_001001409	e-141
DS[96]_34_A_06	Up	3.11	actin, alpha, cardiac muscle 1 like	<i>Danio rerio</i>	NP_001001409	e-148
DS[96]_29_G_10	Up	3.28	actin, alpha, cardiac muscle 1 like	<i>Danio rerio</i>	NP_001001409	e-145
DS[96]_12_G_03	Up	4.58	actin, alpha, cardiac muscle 1 like	<i>Danio rerio</i>	NP_001001409	e-125
DS[96]_21_H_11	Up	6.10	actin, alpha, cardiac muscle 1 like	<i>Danio rerio</i>	NP_001001409	e-127
DS[96]_38_C_06	Up	2.51	actin, beta	<i>Acanthopagrus schlegelii</i>	AAR84618	e-122
DS[96]_05_A_08	Up	1.83	acyl-CoA synthetase long-chain family member 5	<i>Tetraodon nigroviridis</i>	CAG06540	e-102
DS[96]_47_G_04	Up	3.47	aldolase a, fructose-bisphosphate	<i>Danio rerio</i>	NP_919358	e-124
DS[96]_63_E_08	Up	1.86	alpha tubulin, (protein LOC573122)	<i>Danio rerio</i>	NP_001098596	e-120
DS[96]_05_B_06	Up	3.06	amylase-3 protein	<i>Pseudopleuronectes americanus</i>	AAF65827	e-144
DS[96]_70_B_05	Up	3.36	amylase-3 protein	<i>Tetraodon nigroviridis</i>	CAC87127	3.00E-54
DS[96]_34_D_11	Down	4.54	APEX nuclease (apurinic/aprimidinic endonuclease) 2	<i>Xenopus tropicalis</i>	NP_001006804	6.00E-25
DS[96]_01_C_12	Up	1.80	apolipoprotein	<i>Tetraodon nigroviridis</i>	CAG03661	1.00E-38
DS[96]_88_F_11	Up	2.28	apolipoprotein A-I	<i>Danio rerio</i>	NP_571203	1.00E-81
DS[96]_74_G_02	Up	3.34	apolipoprotein A-I	<i>Danio rerio</i>	NP_571203	1.00E-81
DS[96]_72_D_09	Up	7.67	apolipoprotein A-I	<i>Danio rerio</i>	NP_571203	6.00E-73
DS[96]_11_H_09	Up	3.99	apolipoprotein A-I-1 precursor	<i>Oncorhynchus mykiss</i>	O57523	8.00E-76
DS[96]_88_D_02	Up	4.81	apolipoprotein A-I-2 precursor	<i>Oncorhynchus mykiss</i>	O57524	4.00E-71
DS[96]_02_A_11	Up	2.72	apolipoprotein A-IV	<i>Danio rerio</i>	AAH93239	1.00E-73
DS[96]_41_F_02	Up	2.23	apolipoprotein B	<i>Salmo salar</i>	CAA57449	3.00E-24
DS[96]_62_B_05	Up	2.02	apolipoprotein B	<i>Oncorhynchus mykiss</i>	AAG11410	1.00E-19
DS[96]_65_E_08	Up	2.17	apolipoprotein B	<i>Oncorhynchus mykiss</i>	AAG11410	3.00E-19
DS[96]_45_B_03	Up	4.16	apolipoprotein Eb	<i>Danio rerio</i>	NP_571173	2.00E-38
DS[96]_79_H_01	Up	4.42	arachidonate 12-lipoxygenase	<i>Danio rerio</i>	NP_955912	4.00E-23
DS[96]_77_B_08	Up	2.56	arachidonate 12-lipoxygenase	<i>Danio rerio</i>	NP_955912	5.00E-35
DS[96]_86_F_12	Up	3.87	astacin like metallo-protease	<i>Oryzias latipes</i>	NP_001098207	2.00E-83
DS[96]_17_B_07	Up	4.34	astacin like metallo-protease	<i>Oryzias latipes</i>	NP_001098207	7.00E-50
DS[96]_07_A_09	Up	2.17	clq-like protein	<i>Dissostichus mawsoni</i>	ABN45966	3.00E-38
DS[96]_54_A_03	Down	1.69	calcitonin receptor-like receptor	<i>Oncorhynchus gorbuscha</i>	CAD48406	5.00E-56
DS[96]_53_D_10	Down	1.58	calcium binding protein 39	<i>Danio rerio</i>	NP_998666	1.00E-76
DS[96]_71_F_06	Up	2.27	calpain 1 protein	<i>Danio rerio</i>	AAH91999	2.00E-68
DS[96]_37_C_02	Up	1.82	carboxypeptidase H	<i>Paralichthys olivaceus</i>	AAO92752	1.00E-82
DS[96]_27_A_09	Up	2.91	cardiac muscle ATP synthase, alpha 1,	<i>Danio rerio</i>	NP_001070823	7.00E-62
DS[96]_05_C_08	Up	5.76	cell division cycle 14 homolog A	<i>Danio rerio</i>	CAP09233	3.00E-19
DS[96]_30_G_11	Up	2.34	chitin binding Peritrophin-A domain	<i>Danio rerio</i>	AAH45331	4.00E-69
DS[96]_33_H_06	Up	3.75	chitinase	<i>Oncorhynchus mykiss</i>	CAD59687	9.00E-68
DS[96]_69_C_08	Up	4.25	chitinase1	<i>Paralichthys olivaceus</i>	BAD15059	e-127
DS[96]_71_C_06	Up	3.93	chymotrypsinogen 2-like protein	<i>Sparus aurata</i>	AAT45254	1.00E-20
DS[96]_56_B_04	Down	2.01	cofilin 2 (muscle)	<i>Danio rerio</i>	NP_991263	5.00E-84
DS[96]_59_A_01	Down	2.35	cofilin 2 (muscle)	<i>Danio rerio</i>	NP_991263	3.00E-83
DS[96]_67_A_02	Up	5.20	corticotropin-lipotropin A precursor	<i>Oncorhynchus mykiss</i>	Q04617	7.00E-63
DS[96]_26_G_05	Down	1.59	cytochrome P450, family 46, subfamily A, polypeptide 1	<i>Danio rerio</i>	NP_001018358	2.00E-65
DS[96]_11_H_05	Up	1.86	DAZAP2-like protein (deleted in azoospermia-associated)	<i>Takifugu rubripes</i>	NP_001072102	5.00E-59
DS[96]_18_C_06	Down	1.78	dopachrome tautomerase	<i>Salmo salar</i>	ABD73808	1.00E-85
DS[96]_20_B_11	Down	1.66	E3 ubiquitin-protein ligase MARCH2	<i>Danio rerio</i>	Q1LVZ2	2.00E-87
DS[96]_68_G_10	Up	5.08	elastase 2-like protein	<i>Sparus aurata</i>	AAT45251	2.00E-29

# Microarray differentially expressed gene list

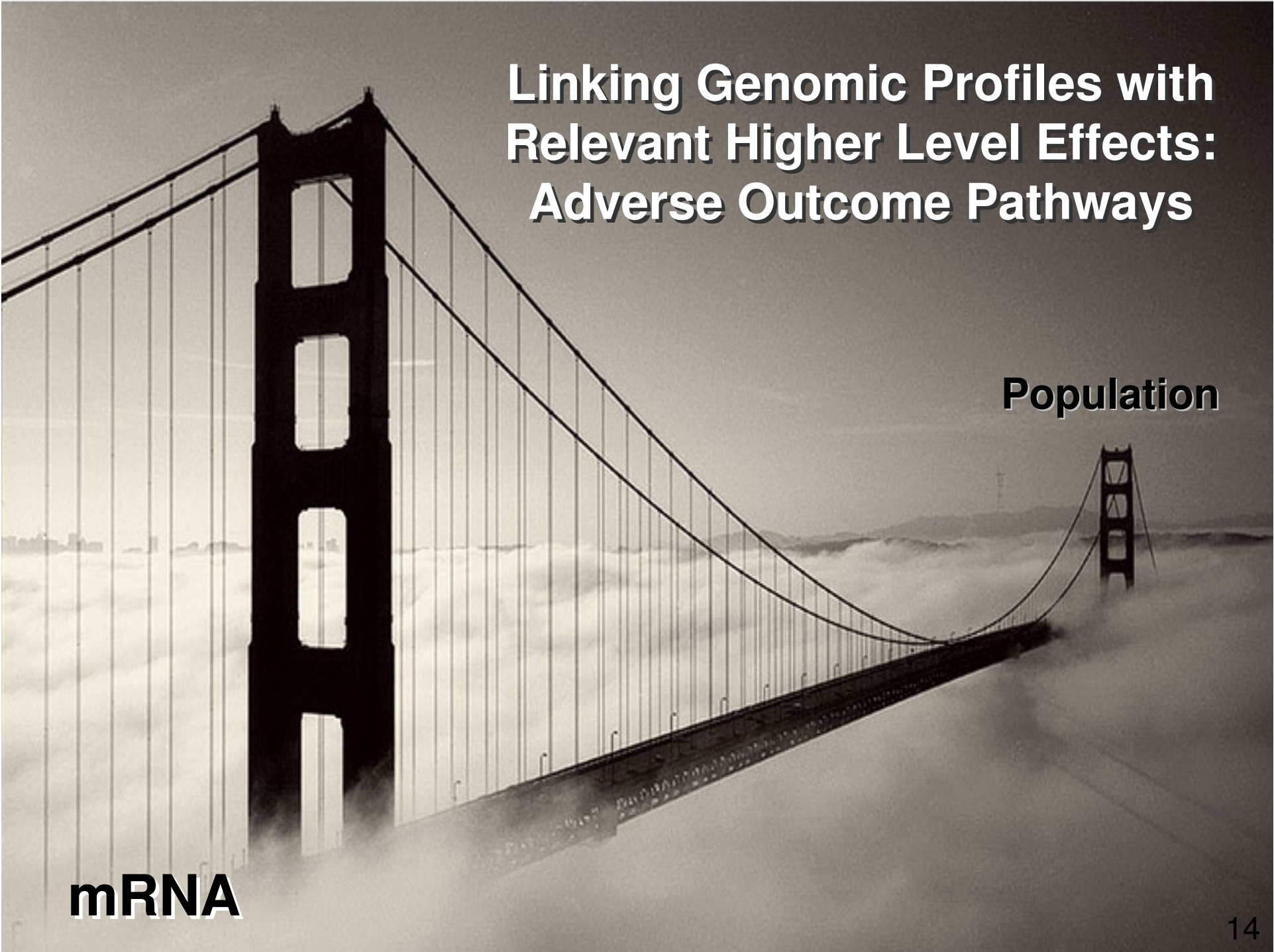
# Microarrays, biomarkers in monitoring



- Provides information on the mode of action
- Allows test organisms to tell us what we should be looking at
- Permits the development and selection of biomarkers without preconceived ideas
- No prior knowledge of the system required
- Identifies specific effects on organism/condition in question





A black and white photograph of the Golden Gate Bridge, viewed from a low angle. The bridge's towers and suspension cables are silhouetted against a bright, hazy sky. The bridge spans across a vast expanse of white clouds that fill the lower two-thirds of the frame. The overall mood is ethereal and atmospheric.

# Linking Genomic Profiles with Relevant Higher Level Effects: Adverse Outcome Pathways

**Population**

**mRNA**

# Linking Molecules

with

# Higher Levels of Biological Organization

...

Bioenergetics

Histopathology

Immunology

Physiology

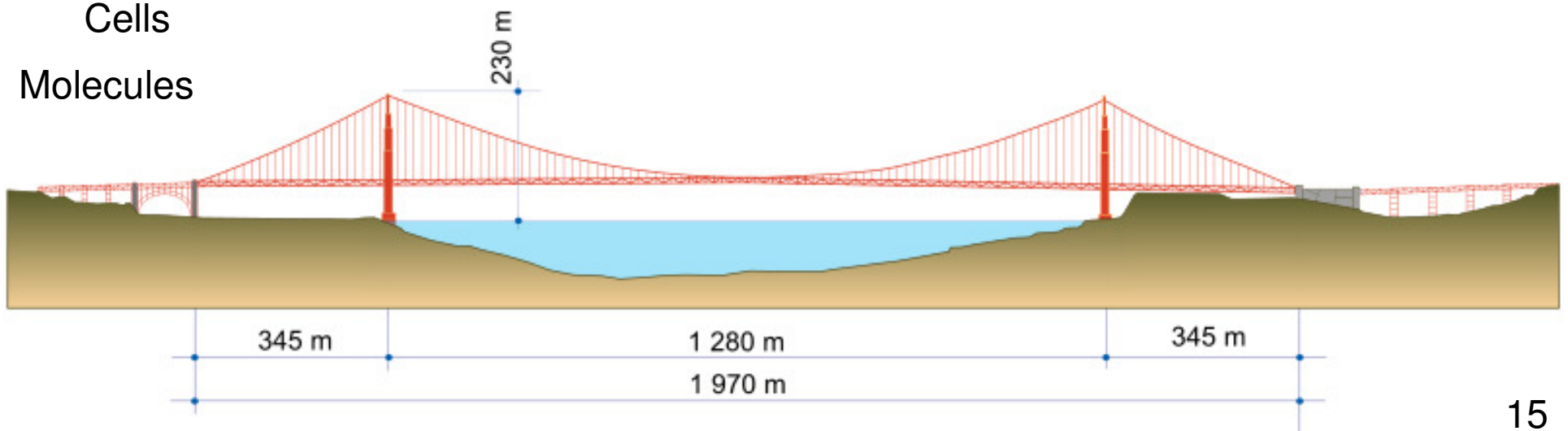
Detoxification

Cells

Molecules

Development and  
Reproduction

Population and  
Community



**Linking  
Molecules**

**with**

**Higher Levels of Biological  
Organization**

...

Bioenergetics

Histopathology

Immunology

Physiology

Detoxification

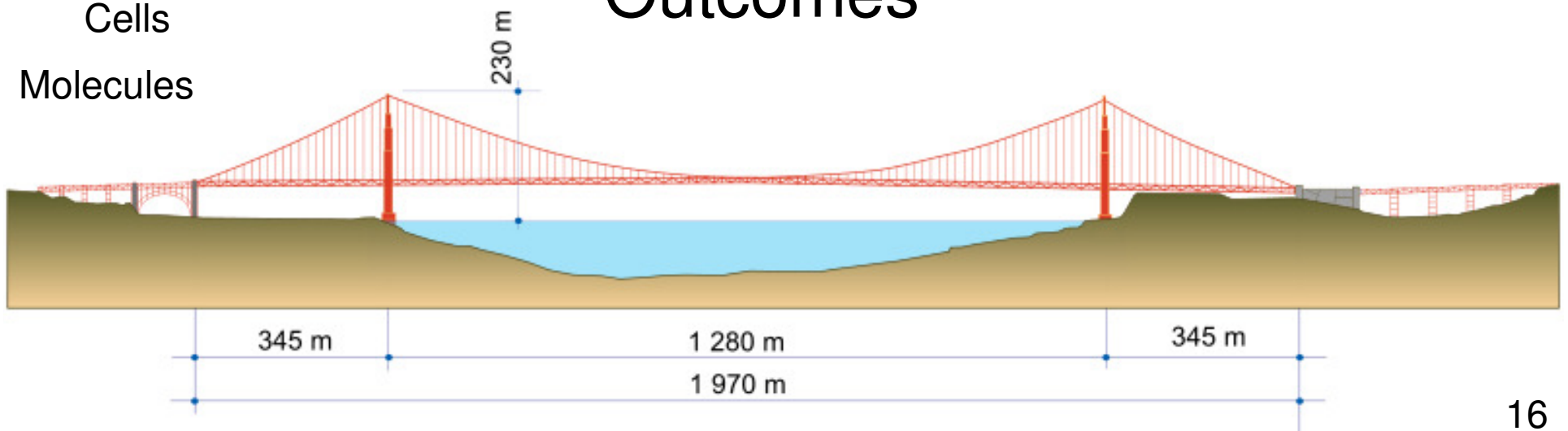
Cells


Molecules

Development and  
Reproduction

Population and  
Community

**Adverse  
Outcomes**

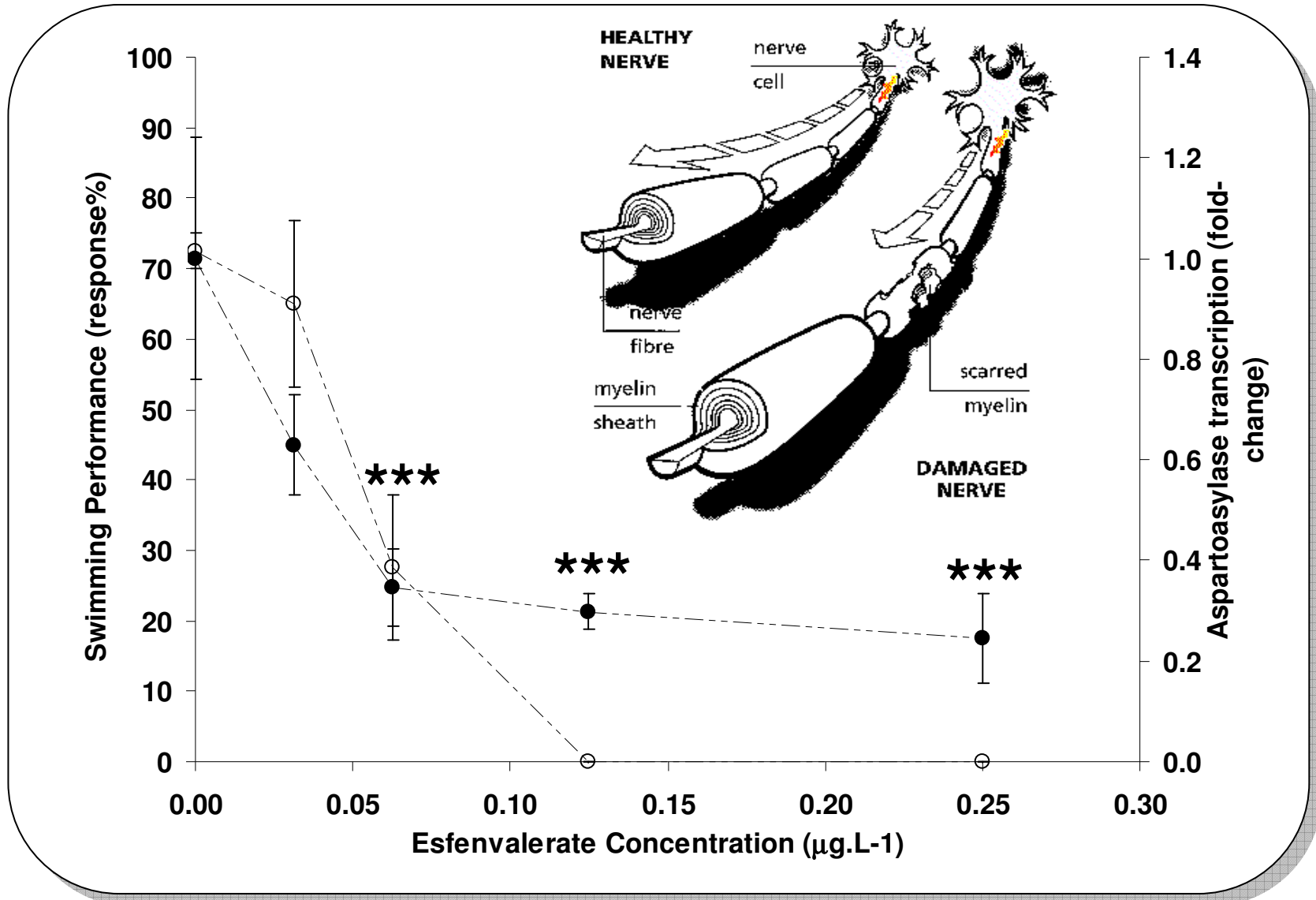




**Measure it**  
**Swim it**  
**...**  
**Bridge it**



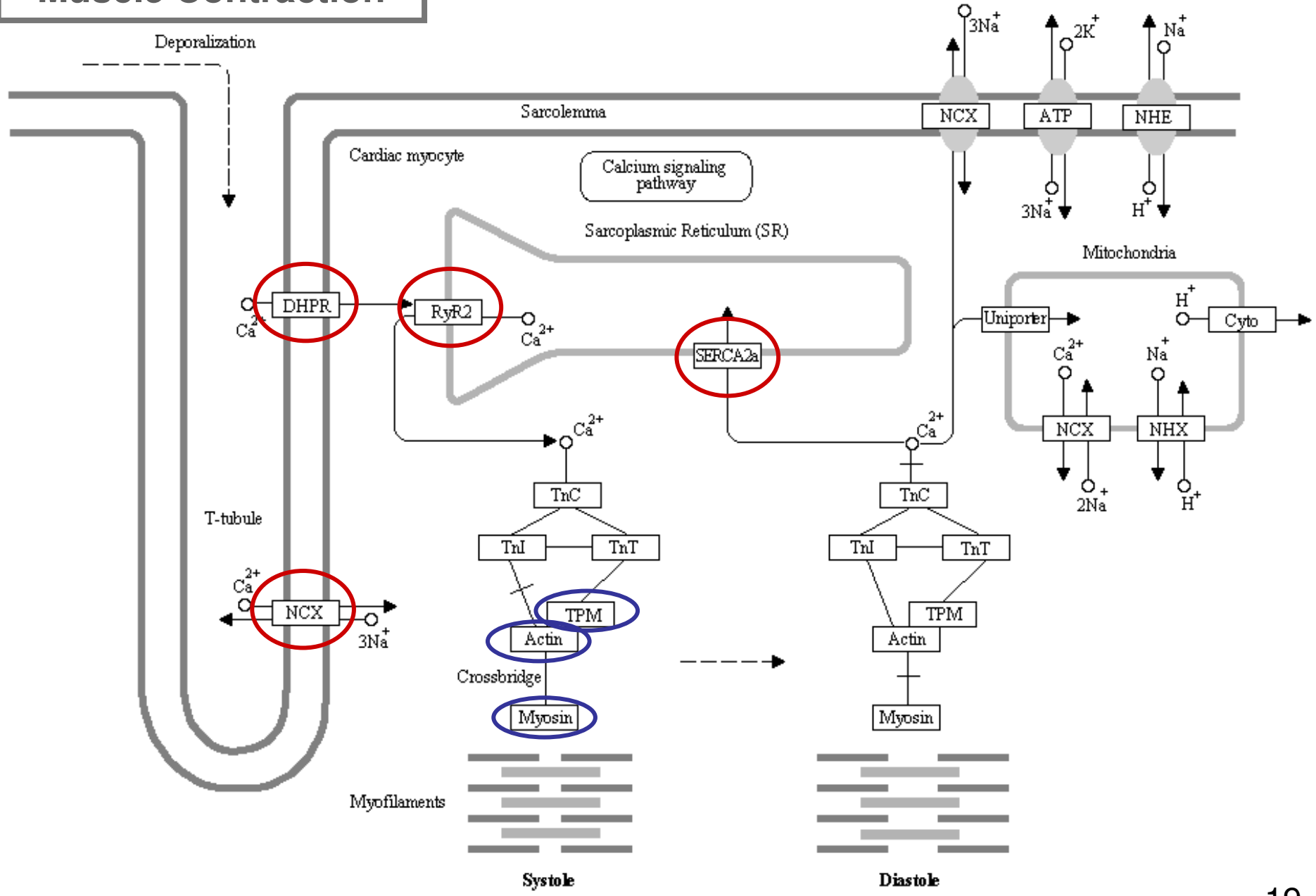
Connon RE, et al (2009). Linking mechanistic and behavioral responses to sublethal esfenvalerate exposure in the endangered delta smelt; *Hypomesus transpacificus* (Fam. Osmeridae). BMC Genomics.

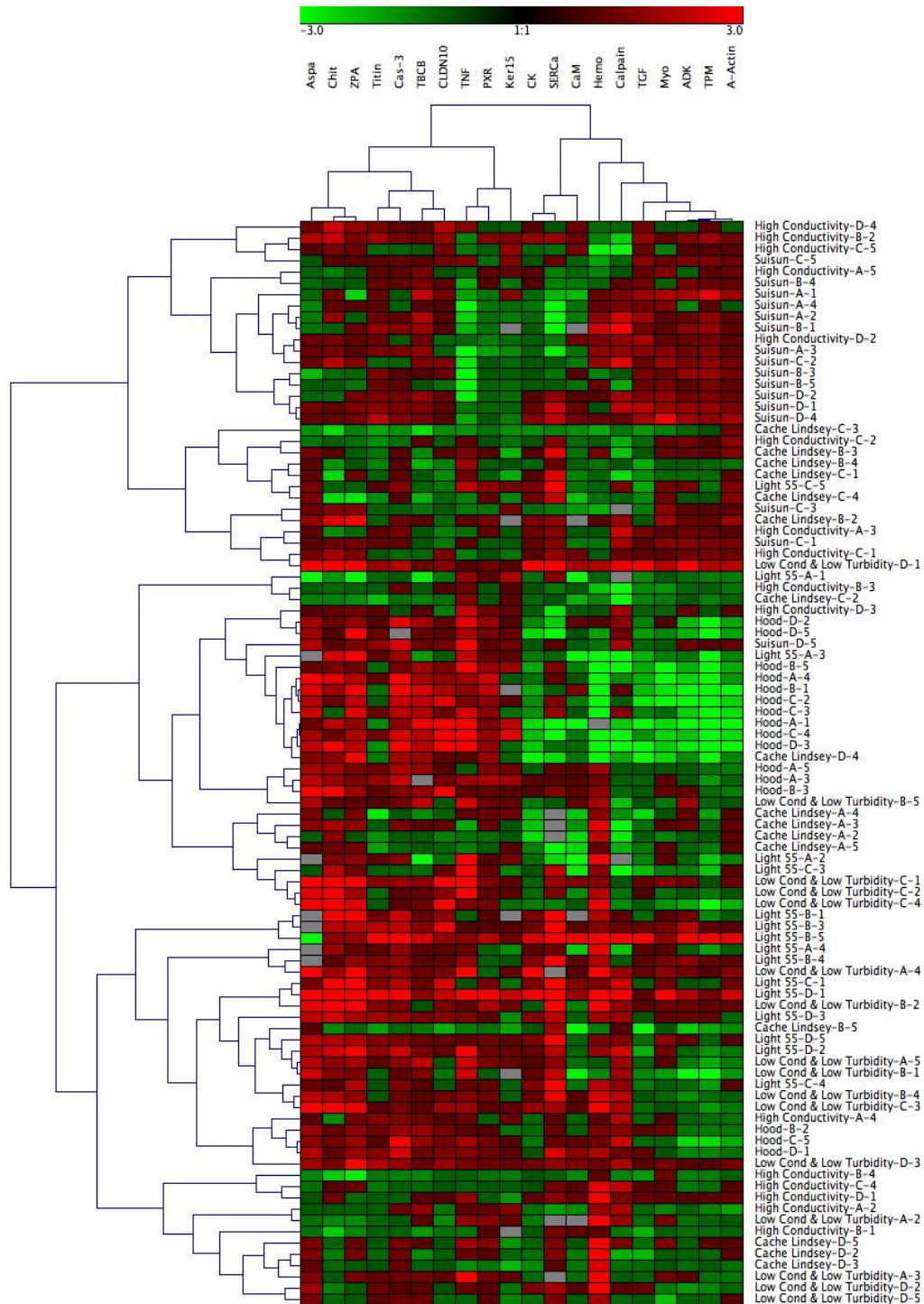


Swimming performance and aspartoacylase transcription in larval delta smelt exposed to esfenvalerate



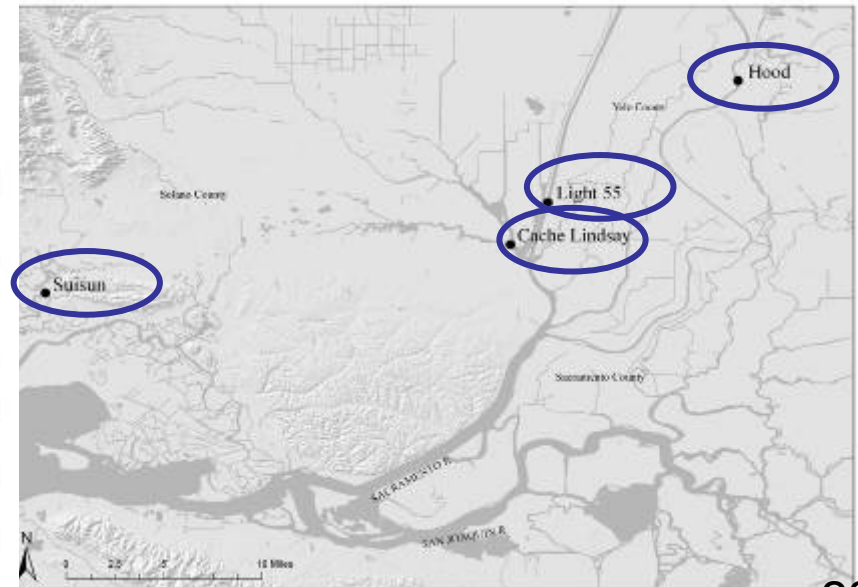
# Muscle Contraction





## Genomic profiling:

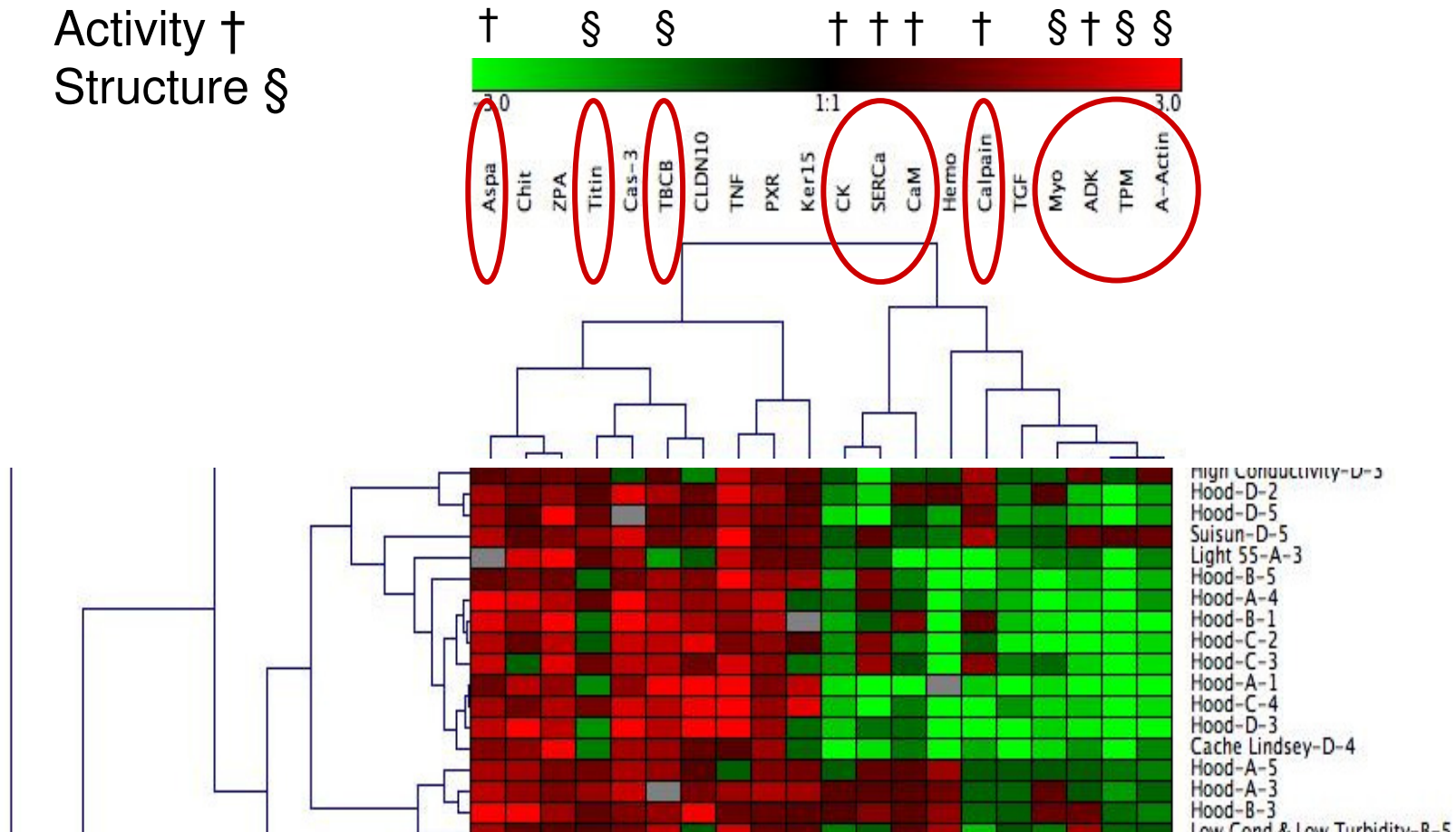
Responses in juvenile delta smelt exposed for 7 days to ambient samples from four sites in the Bay-Delta System, along with respective high and low EC controls



# Genomic profiling: Hood

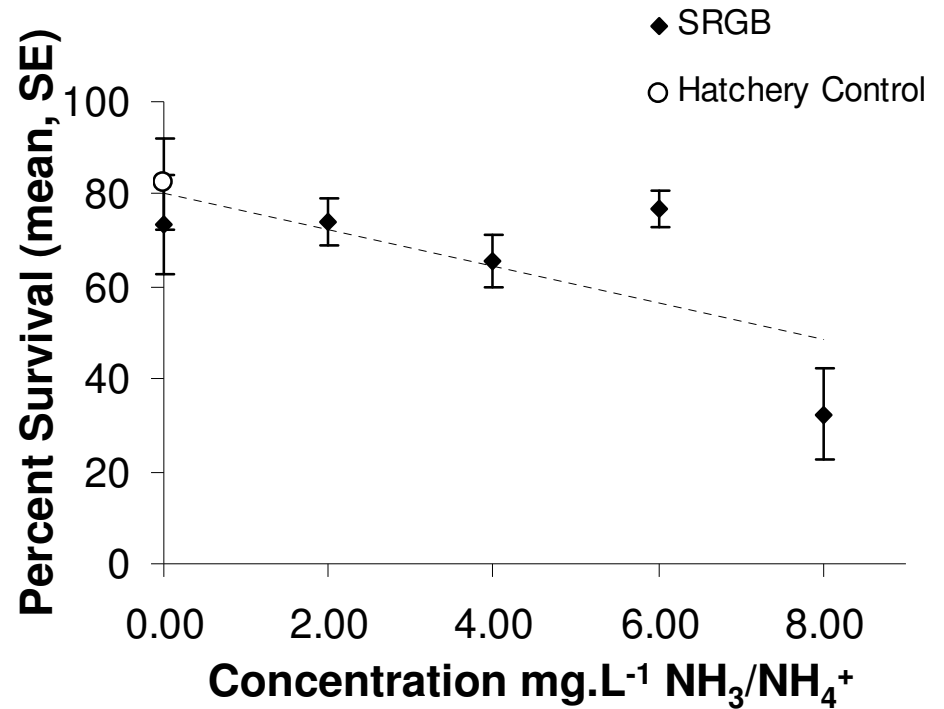
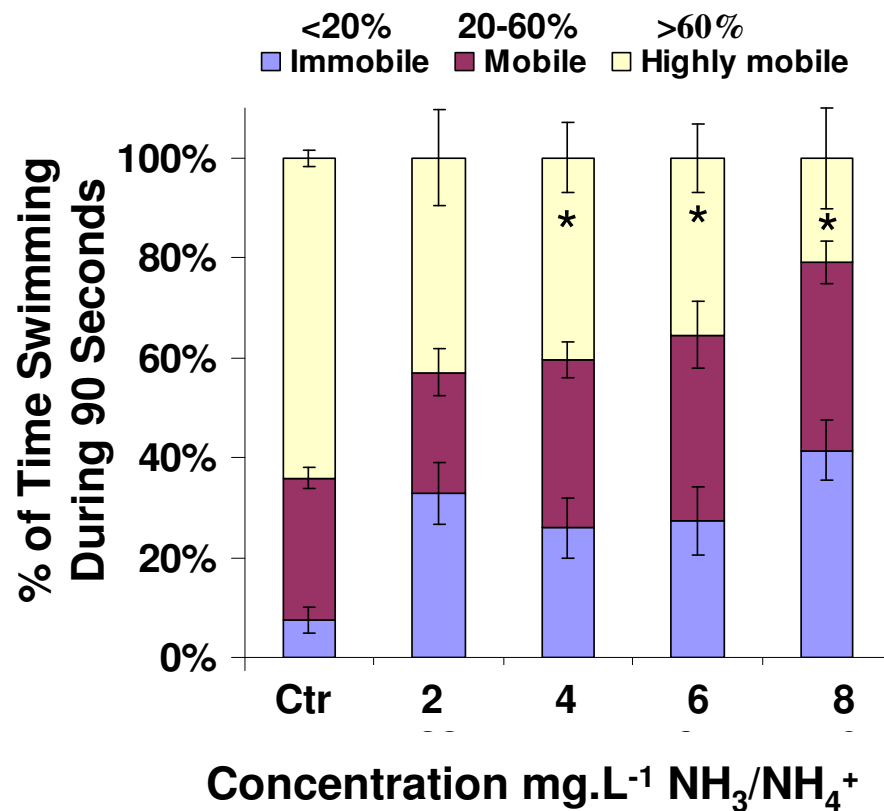
Key:

Activity †  
Structure §



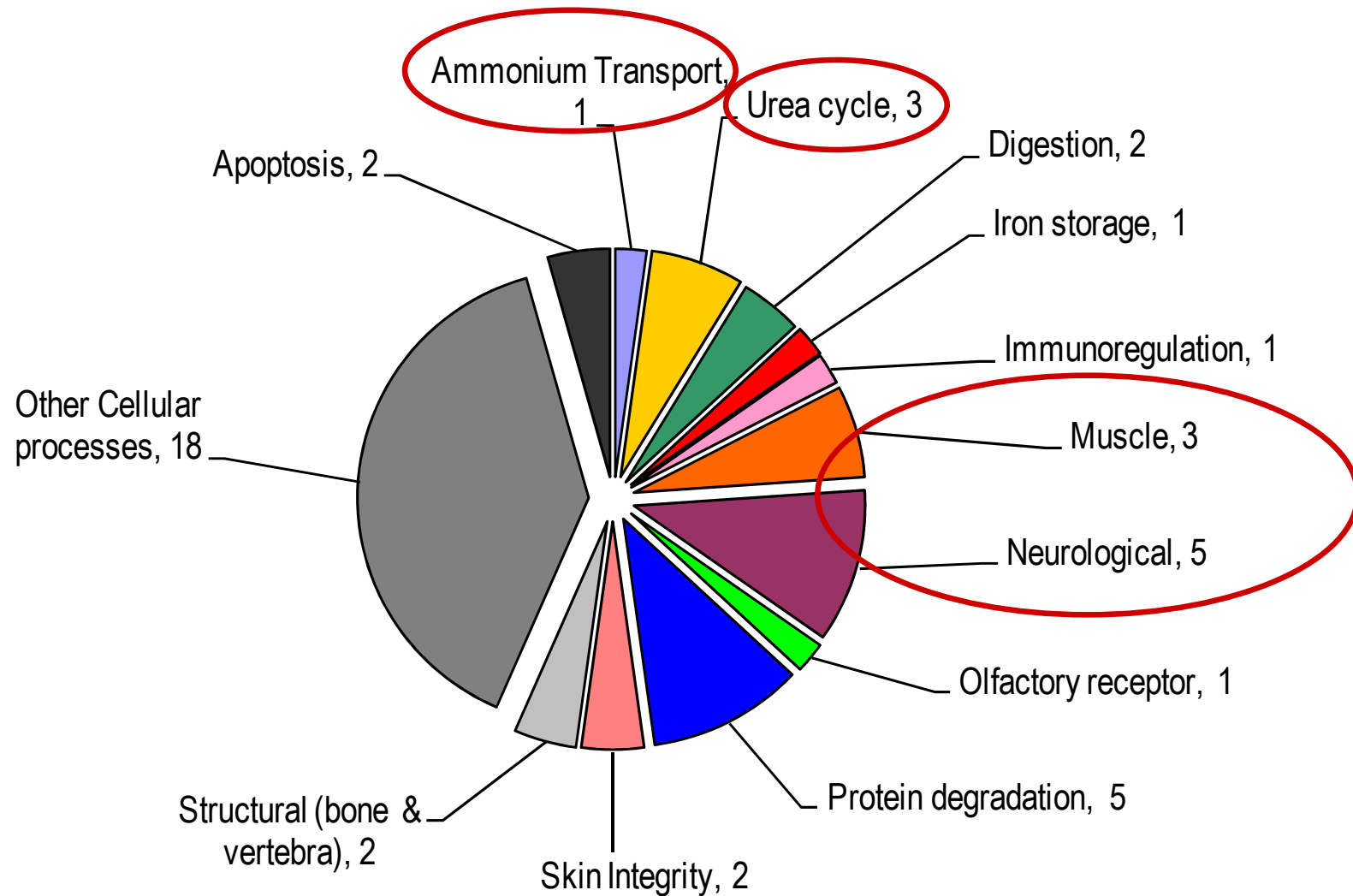
# Survival and swimming of Larval Delta Smelt after 7-d Exposure to **Ammonium Chloride**

(in Sacramento River water from Garcia Bend)



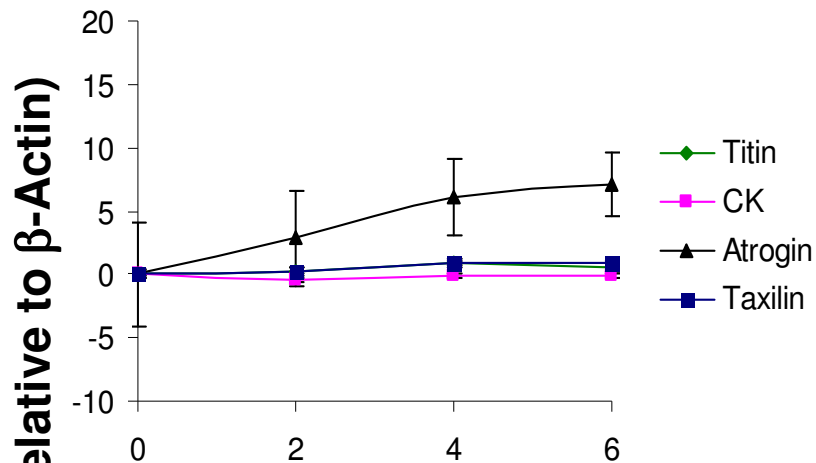
	<b>NH<sub>4</sub>+Cl</b> <b>(mg.L-1 NH<sub>3</sub>/NH<sub>4</sub><sup>+</sup>)</b>
<b>7-d LC50</b>	<b>7.5</b>

# Functional classification of genes responding to 4 mg.L<sup>-1</sup> NH<sub>3</sub>/NH<sub>4</sub><sup>+</sup> from NH<sub>4</sub>Cl

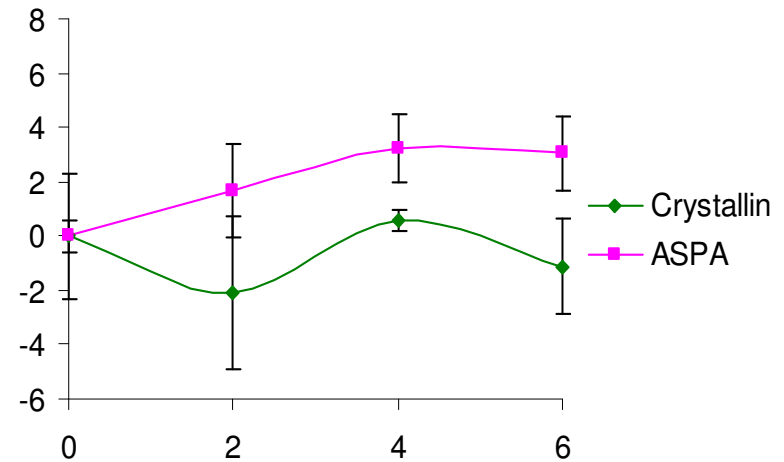




### Muscular responses

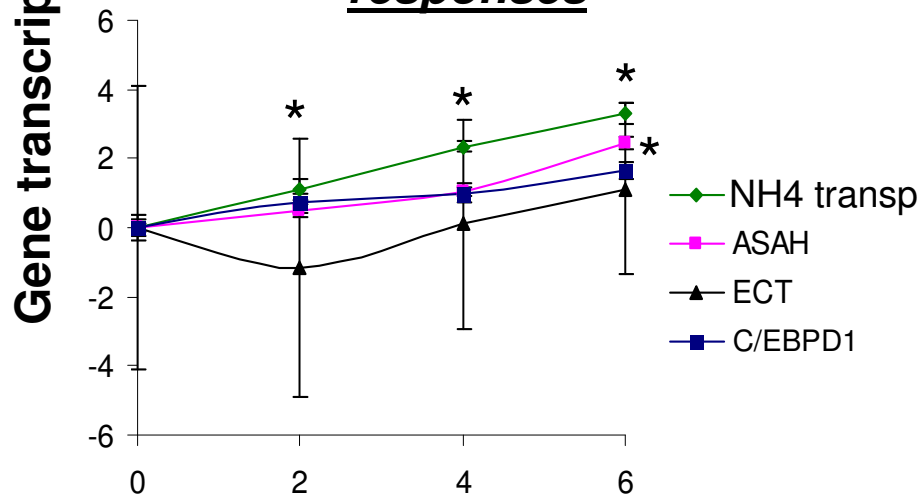


### Neurological responses



Ammonia/ $\mu$ m Concentration ( $\text{mg.L}^{-1}$ )

### Direct link to ammonia responses

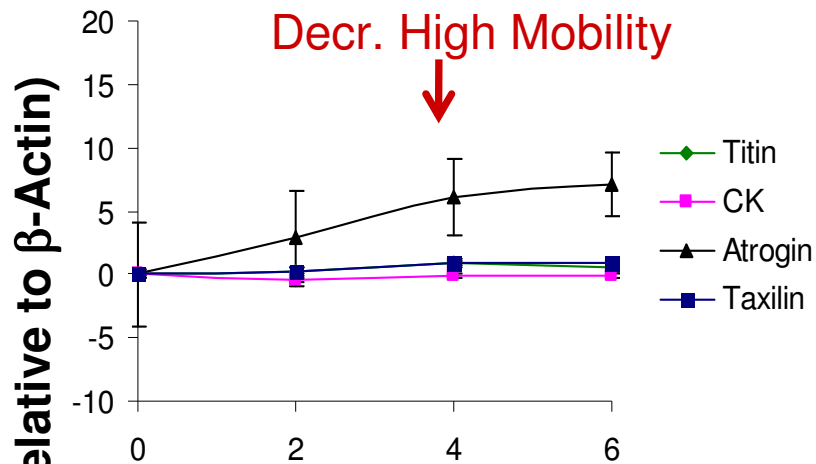


Ammonia/ $\mu$ m Concentration ( $\text{mg.L}^{-1}$ )

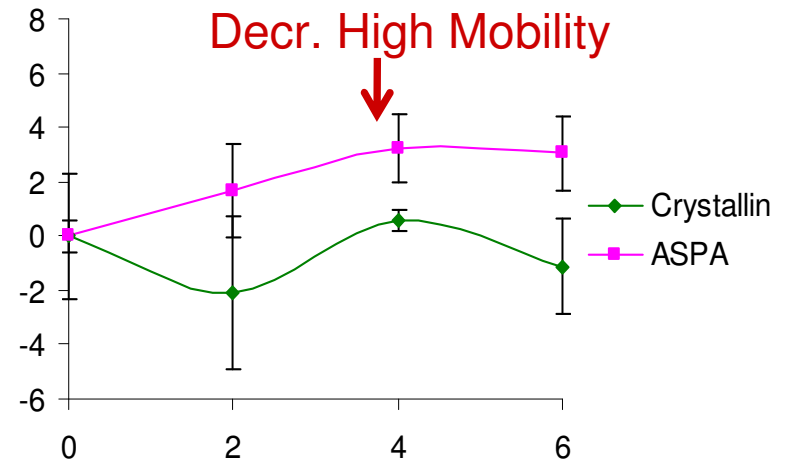
### Molecular biomarker responses (n=10)

Genes were selected from top 10 statistically significant responses identified with microarray – and others from previous tests (e.g. Creatine Kinase, Aspartoacylase)

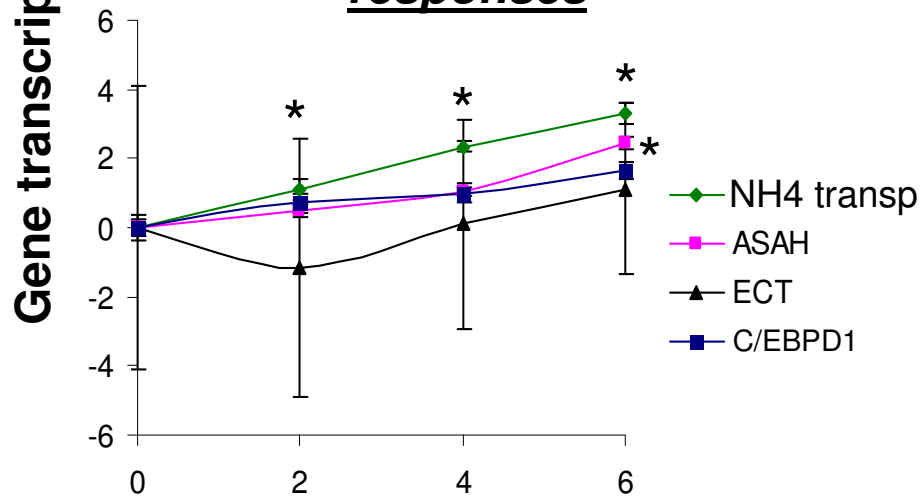
### Muscular responses



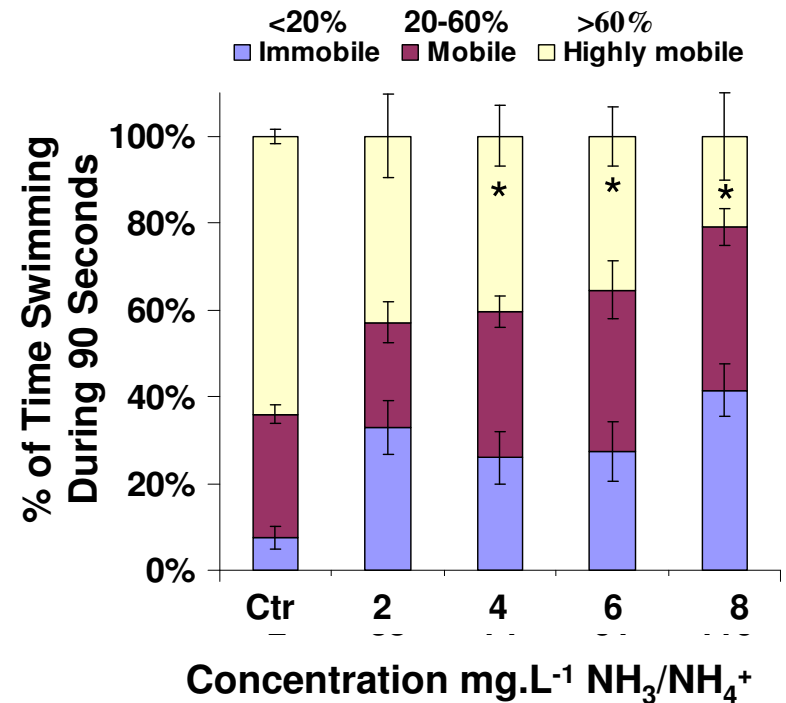
### Neurological responses



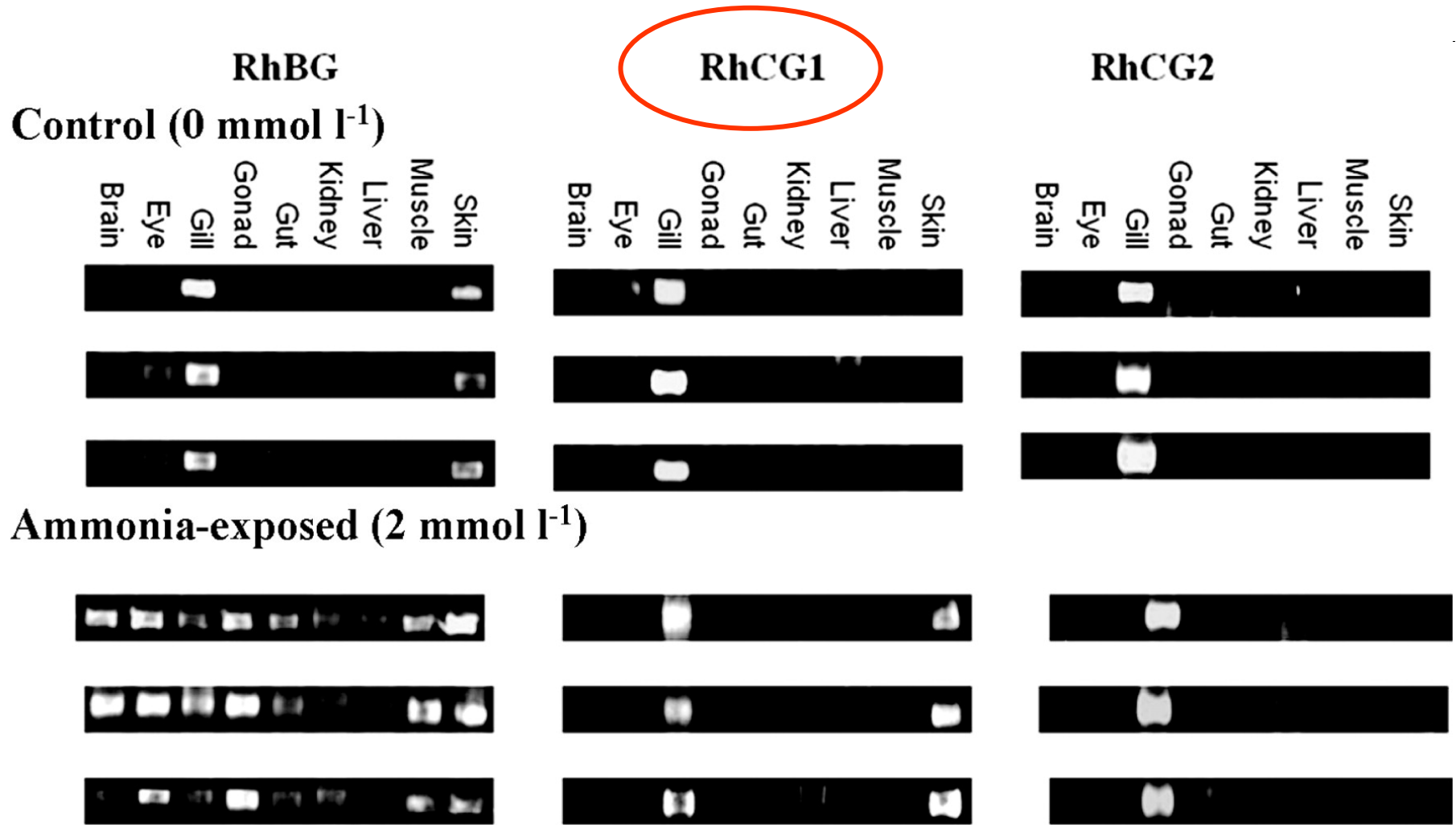
### Direct link to ammonia responses



Ammonia/ $\mu$ m Concentration ( $\text{mg}\cdot\text{L}^{-1}$ )



Hung CYC, et al (2007) Rhesus glycoprotein gene expression in the mangrove killifish *Kryptolebias marmoratus* exposed to elevated environmental ammonia levels and air. J. Experimental Biology.



Quantitative PCR mRNA expression of *RHbG*, *RhCG1* and *RhCG2* in control and ammonia-exposed *K. marmoratus*.



# Current, future and proposed work

- Generating contaminant-specific genomic profile database, using microarray and qPCR technology
- Assaying field based exposures
- Developing biomarkers linked to reproduction

Questions we're seeking answers to:

- Can we use profiling system to aid TIE investigations based on chemical modes of action?
- Do all fish species respond comparatively to equitoxic contaminant concentrations?

Molecular Biomarkers as Tools in  
Toxicity Testing and Field  
Monitoring  
?

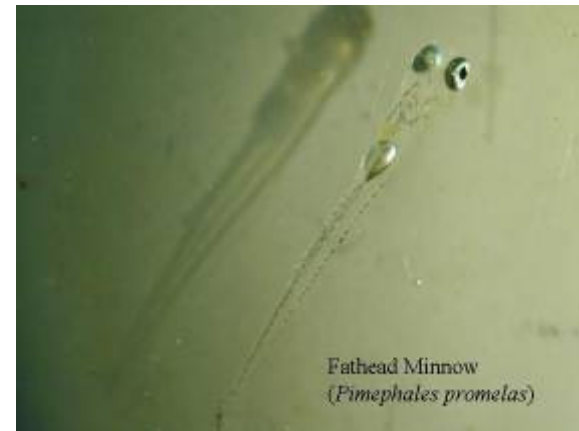


## Major Challenges:

- ❑ Laboratory toxicity testing with resident species;
  - Often difficult to culture and handle.
  - Require extensive method development.
  - Can generally not be used for in-depth studies evaluating ecologically important chronic endpoints, such as reproductive success, behavior and/or growth.
  
- ❑ Interpretation of biomarker data collected in field monitoring studies.

# Model Fish Species in Toxicity Testing

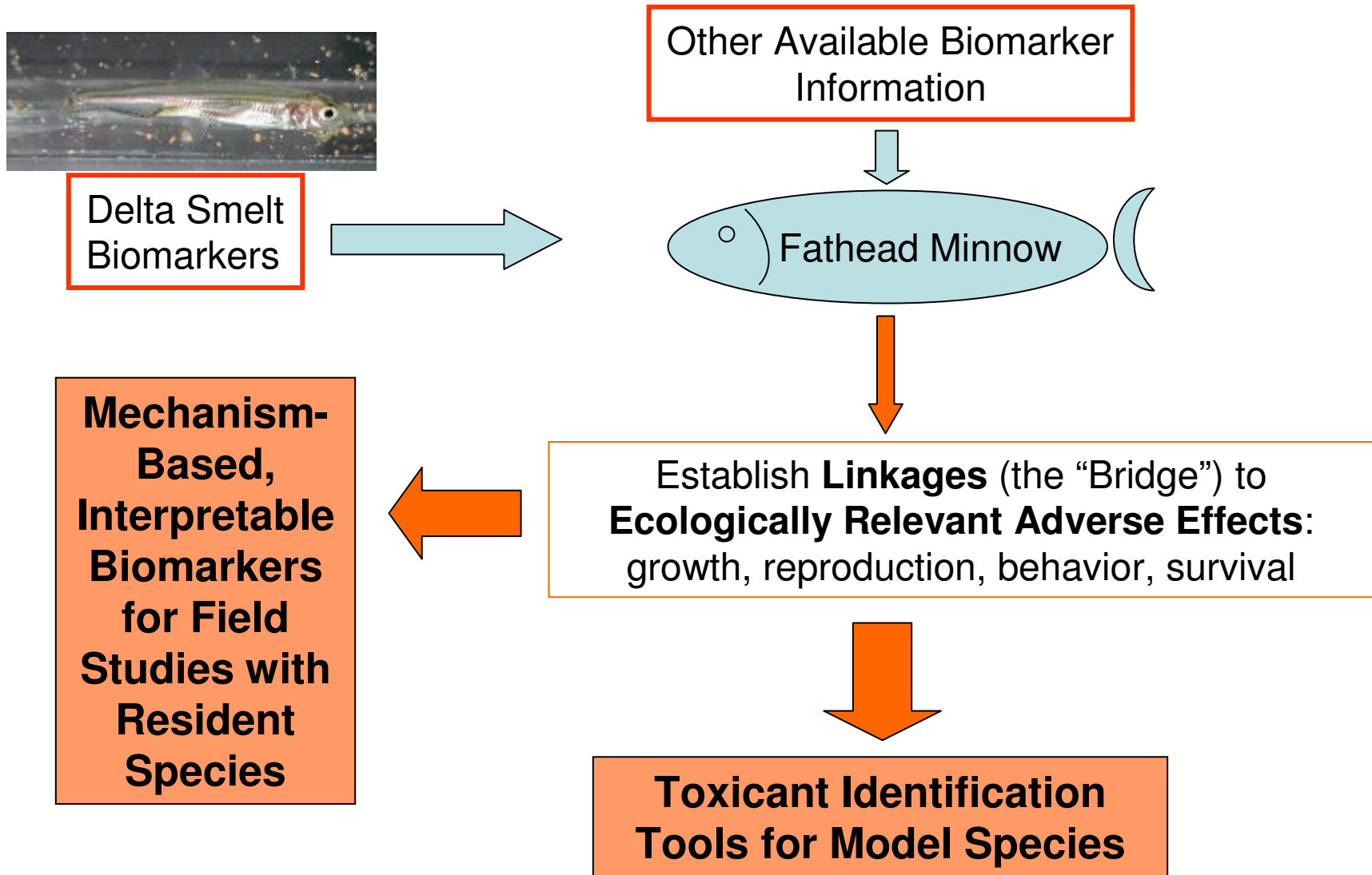
- **Rainbow Trout** (*survival, embryo test, growth*)
- **Fathead Minnow** (*growth, survival, reproduction*)
- **Zebrafish** (*everything?*)



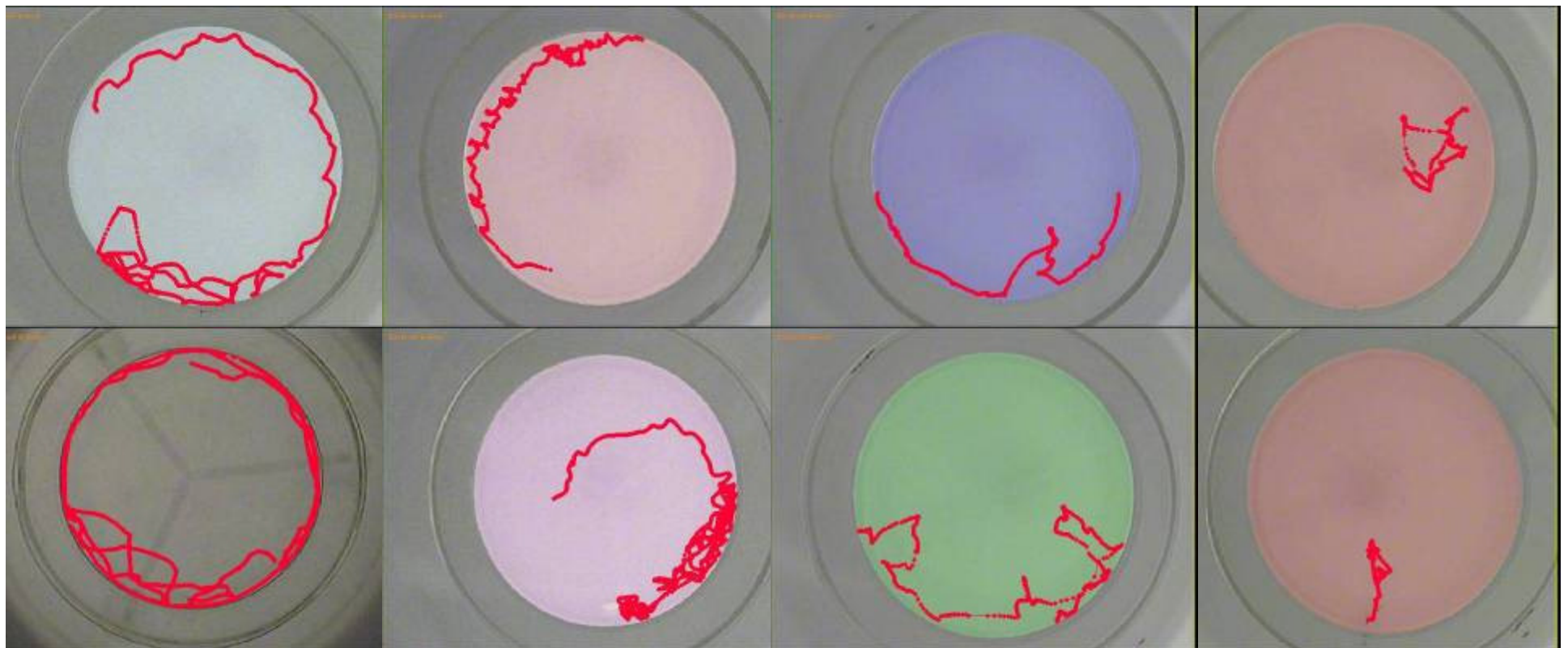
# Important Questions:

- **Are expression patterns of selected biomarker genes reflective of species sensitivity and stressor type across species?**
- **Are molecular biomarkers mechanistically linked to contaminant type and ecologically relevant effects?**

# ..... Connecting the Dots.....



# Neuromuscular Effects of Pesticides: Swimming Behavior of Larval Fathead Minnow after 24 h Exposure to Talstar® (Bifenthrin)



Control

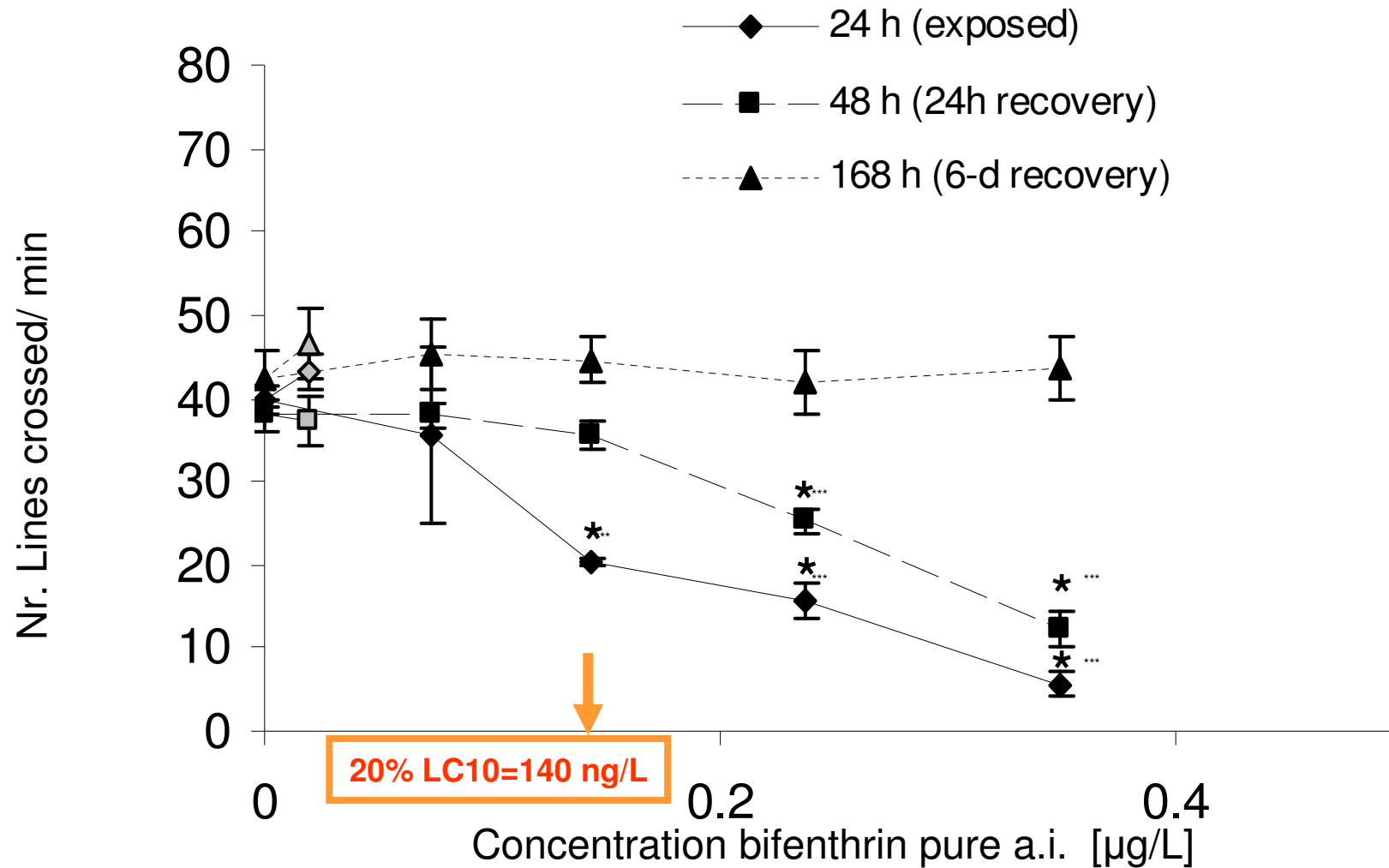
20% LC10

33% LC10

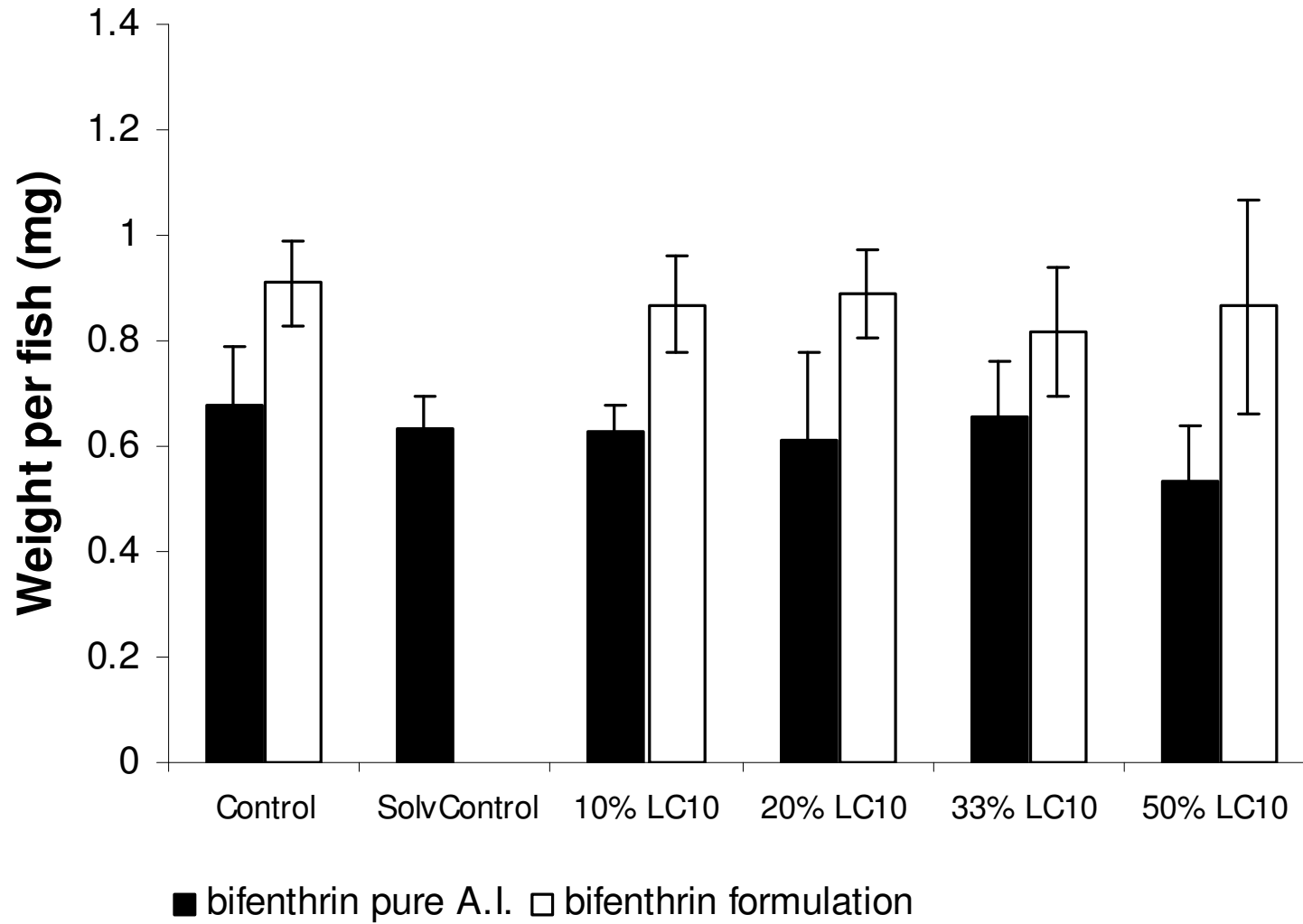
50% LC10



# Swimming Activity in Fathead Minnows Exposed to Bifenthrin for 24 h

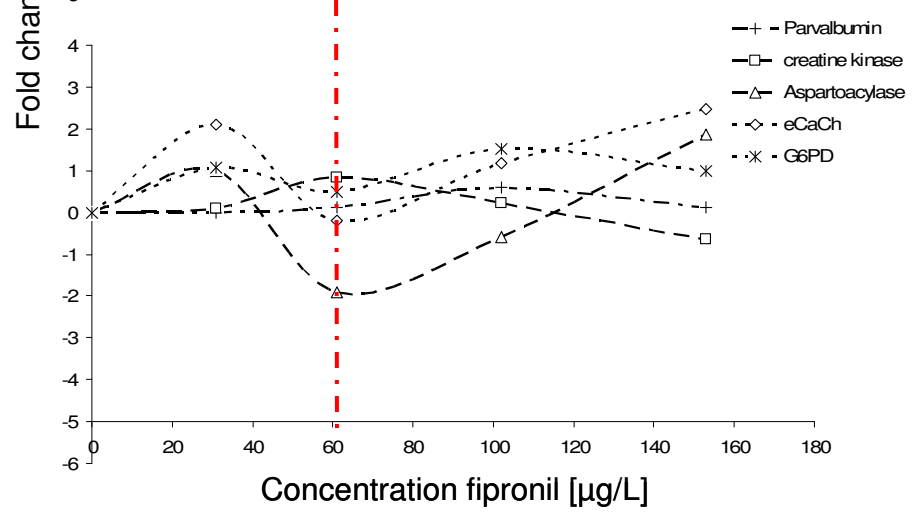
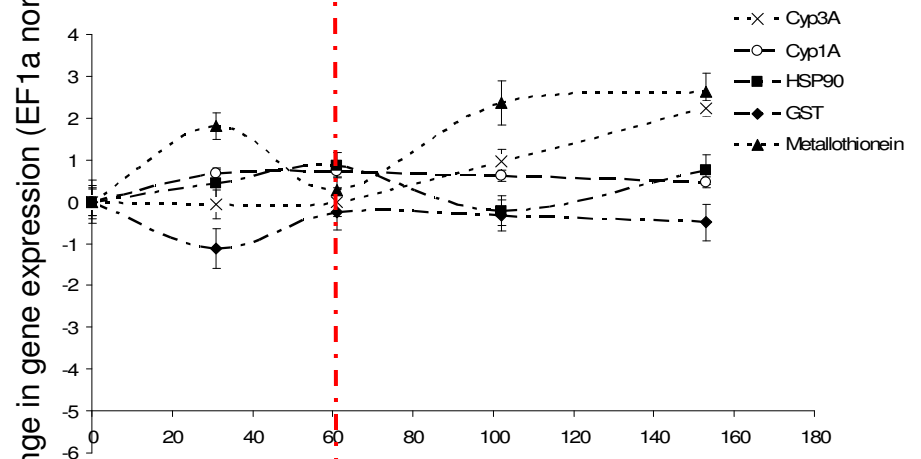
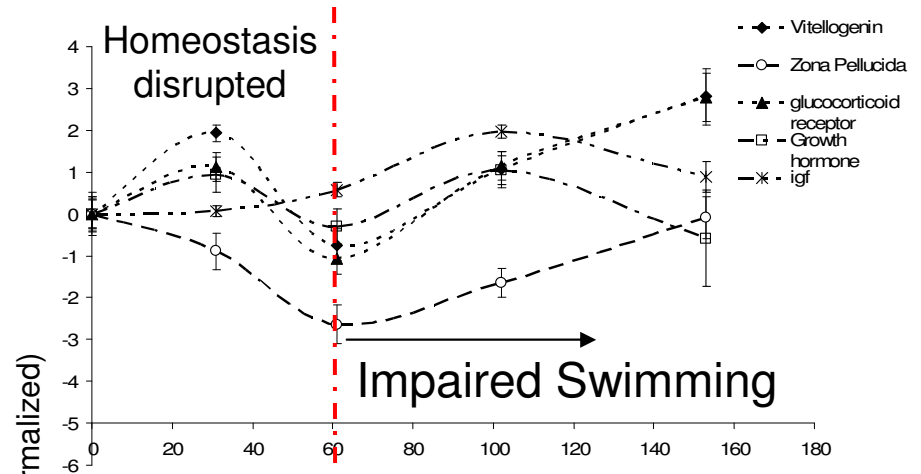
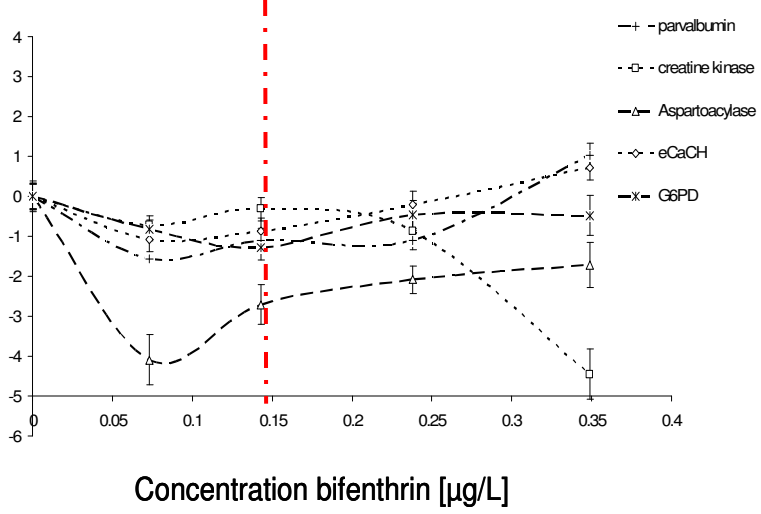
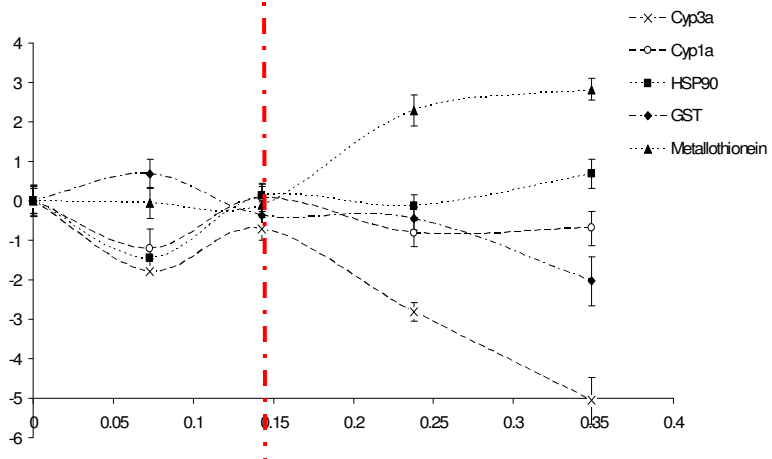
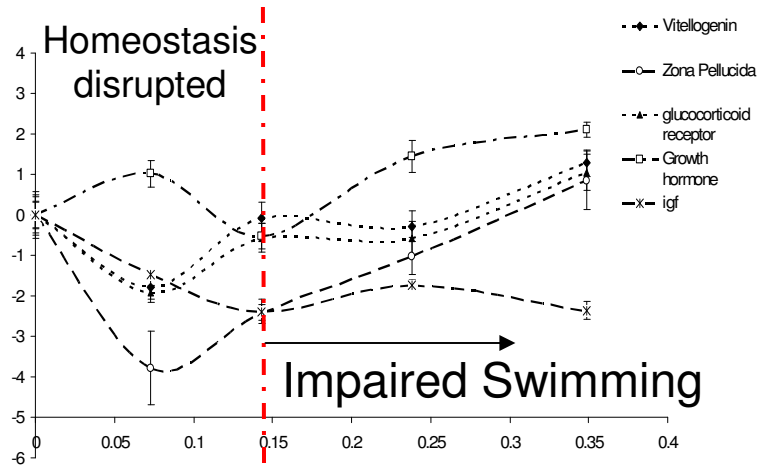


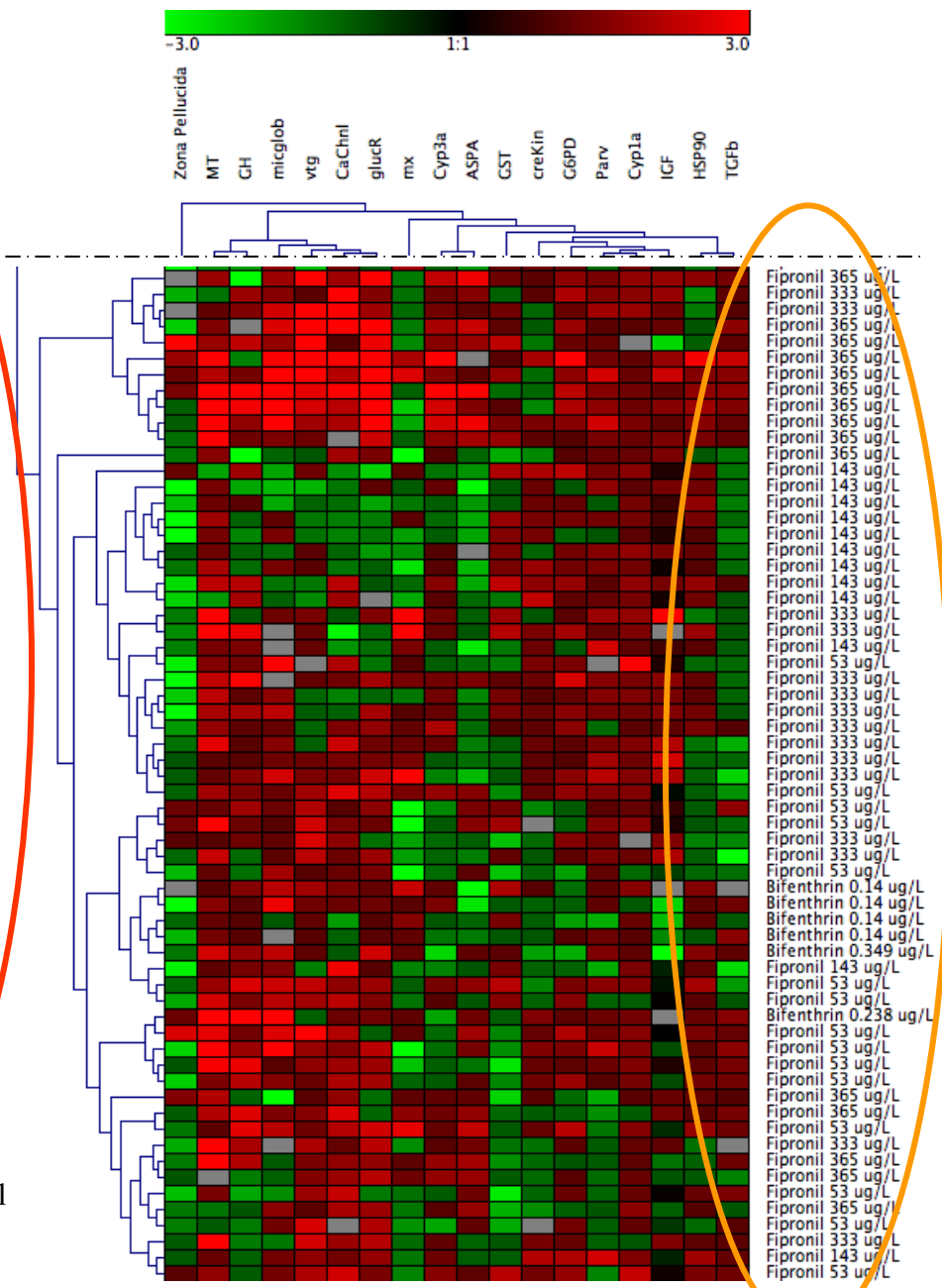
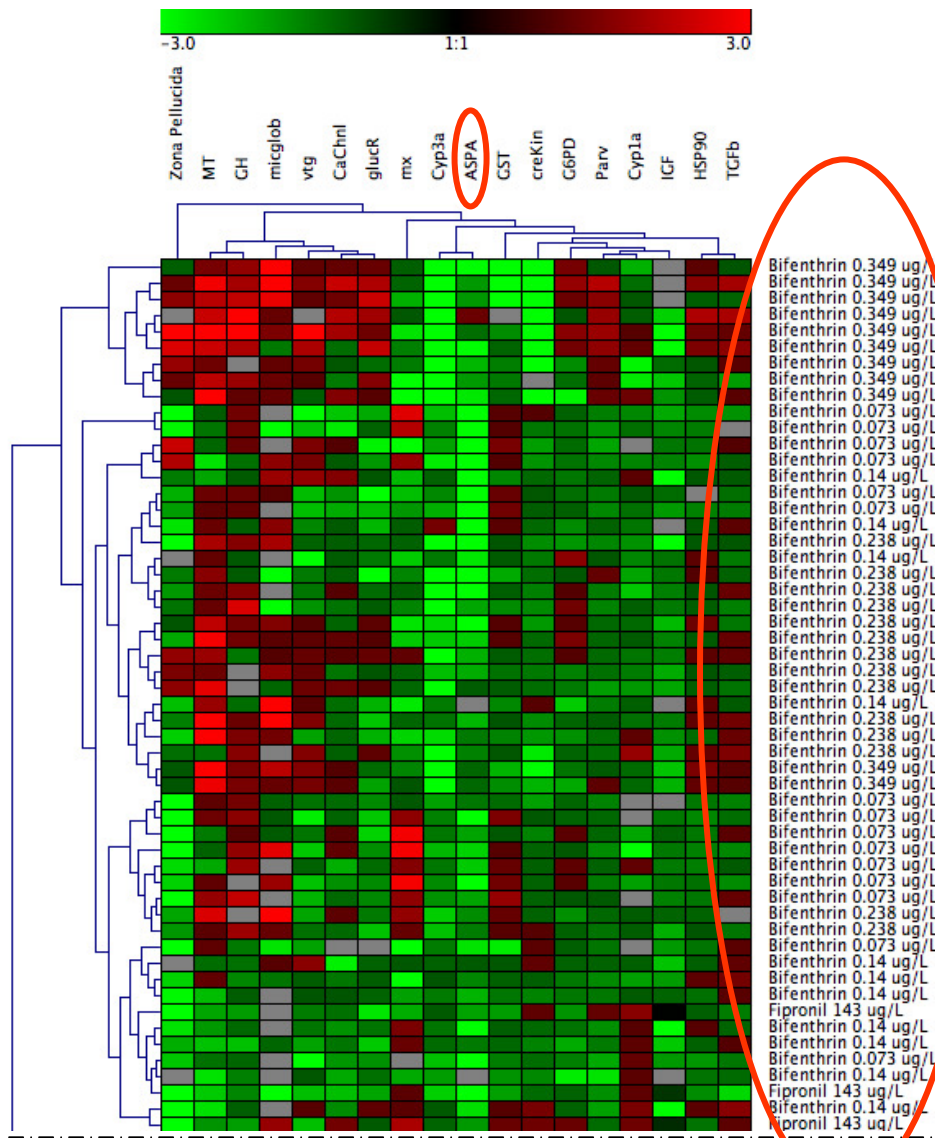
# Growth in Fathead Minnows



# Selected Molecular Biomarkers in Fathead Minnow

<b>Gene Function</b>	<b>Gene</b>
<b>General Stress Response</b>	HSP70, HSP90
<b>Detoxification</b>	Metallothionein, Glutathione-S Transferase, CYP1A
<b>Endocrine Disruption/ Development</b>	Vitellogenin, CYP3A, Zona Pellucida Protein, Glucocorticoid Receptor
<b>Iron Storage</b>	Ferritin
<b>Neuromuscular Function/Energy</b>	Parvalbumin, Creatine Kinase, Epithelial Ca <sup>+</sup> Channel, Aspartoacylase, Titin, Myozenin, G6PD
<b>Growth</b>	Insuline-Like Growth Factor, Growth Hormone
<b>Apoptosis</b>	Caspase 1
<b>Immune System</b>	Microglobulin, IL8, Mx Protein, Nramp





**Genomic fingerprinting (Heatmaps).**

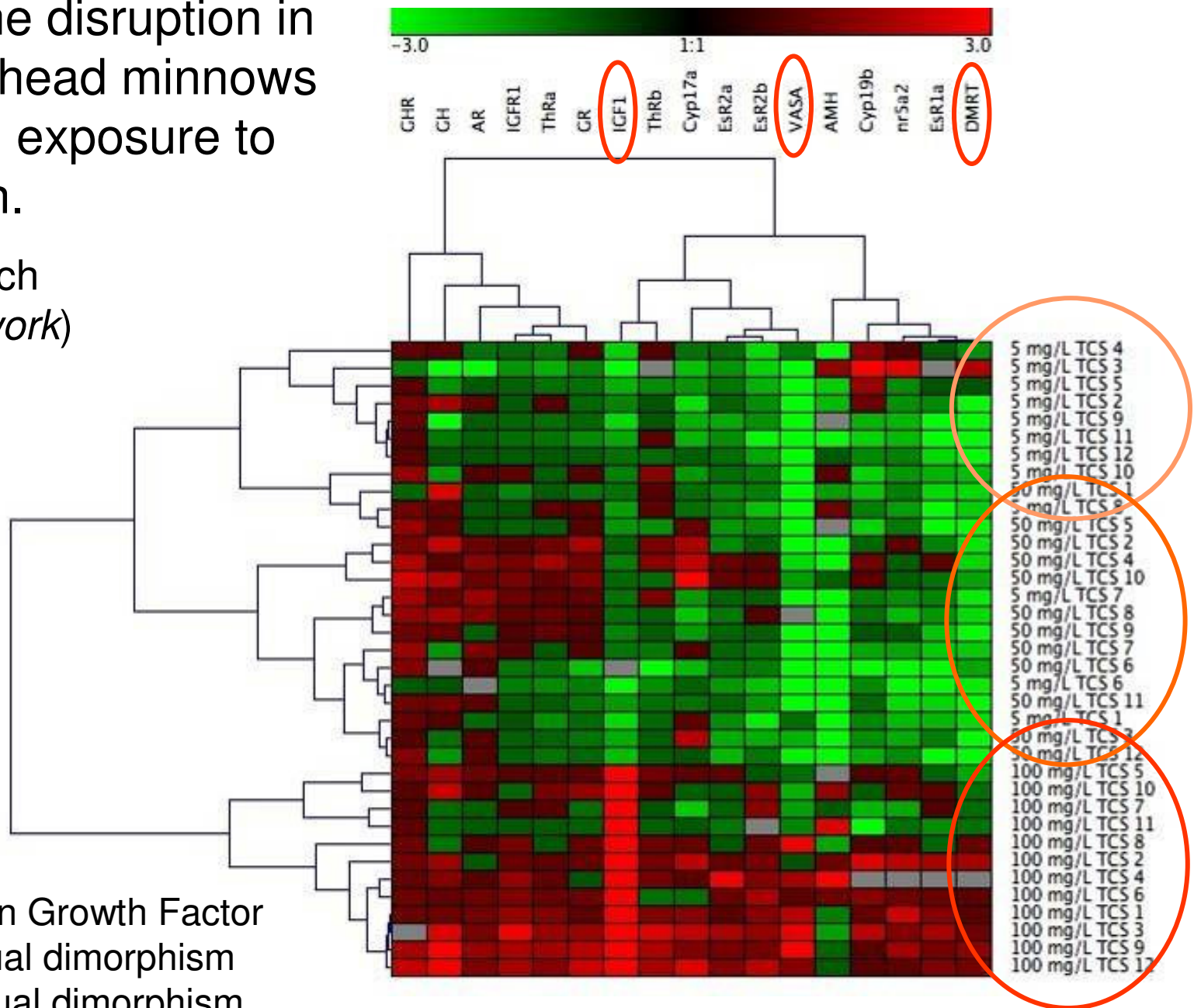
Molecular biomarkers distinguish between bifenthrin and fipronil exposed fathead minnow.

Beggel S, Werner I, Cannon RE, Geist JP. (in preparation). Changes in gene expression in larval fathead minnow (*Pimephales promelas*) following short-term exposure to bifenthrin and fipronil.



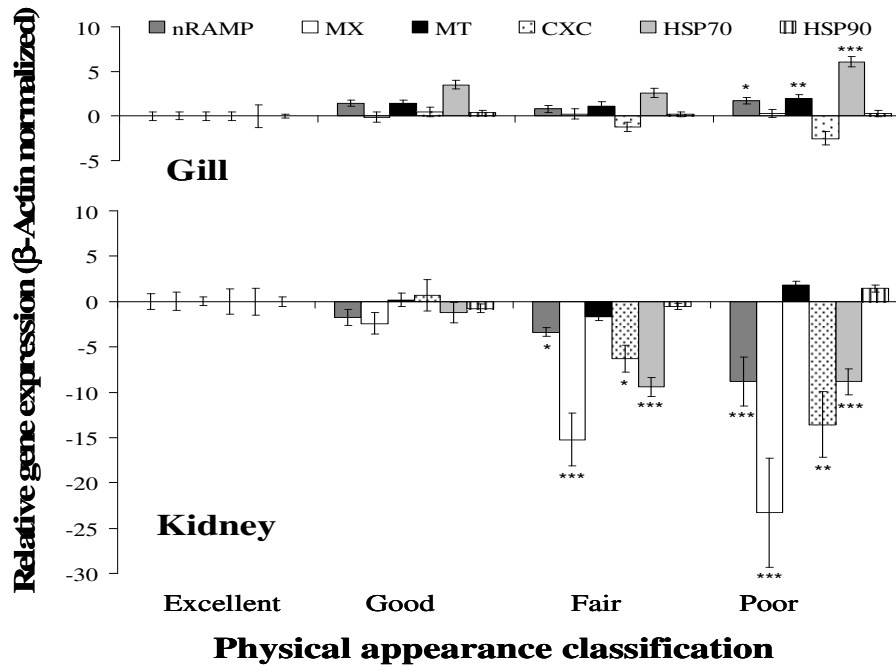
Endocrine disruption in larval fathead minnows following exposure to Triclosan.

Erika Fritsch  
(*Current work*)

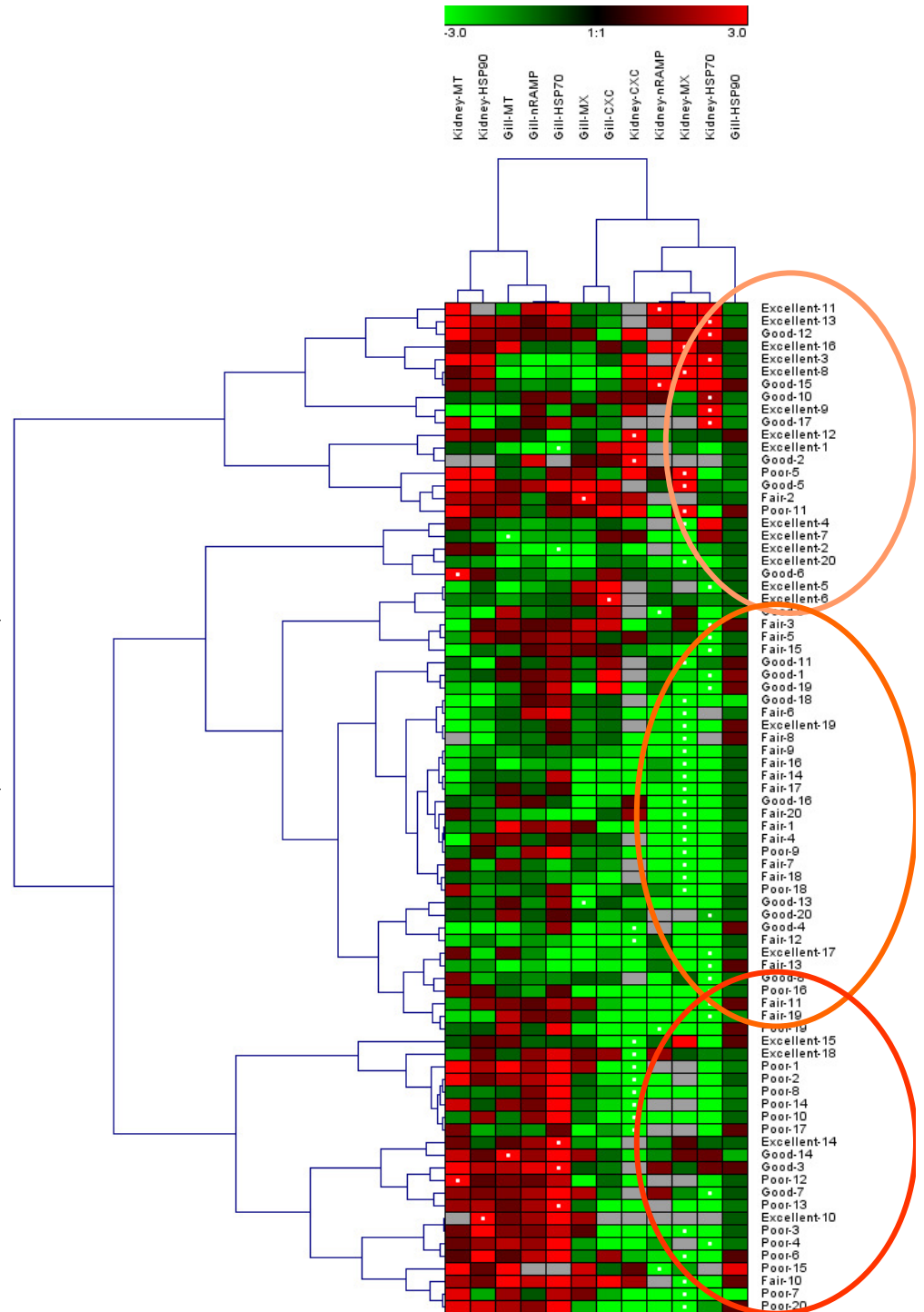


IGF1=Insulin Growth Factor  
VASA: sexual dimorphism  
DMRT: sexual dimorphism

# Expression of selected biomarker genes in Columbia River Steelhead Trout



*D'Abronzio et al., in prep.*



# Conclusions

- Microarray technology allows us to identify new mechanism-based molecular biomarkers.
- The multi-biomarker approach (heatmaps) integrates old and new information.
- Shows great promise in distinguishing the impact of different stressors (TIE).
- Allows mechanism based interpretation of biomarker signals.
- Leads to a better understanding of linkages (“adverse outcome pathways”).





# Acknowledgments

- ❖ The Interagency Ecological Program, Sacramento, CA, for funding the microarray and biomarker work.
- ❖ The UC Davis Fish Conservation and Culture Laboratory, Byron, CA, for providing delta smelt.
- ❖ The staff of the UCD Aquatic Toxicology Lab.
- ❖ Sebastian Beggel & Erika Frisch, Graduate Students in I. Werner's lab.